# Early Vision and Visual System Development

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## Studying the visual system (1)

The visual system can be (and is) studied using many different techniques. In this course we will consider:

**Psychophysics** What is the level of human visual performance under various different conditions?

**Anatomy** Where are the visual system parts located, and what do they look like?

Gross anatomy What do the visual system organs and tissues look like, and how are they connected?

**Histology** What cellular and subcellular structures can be seen under a microscope?

# Studying the visual system (2)

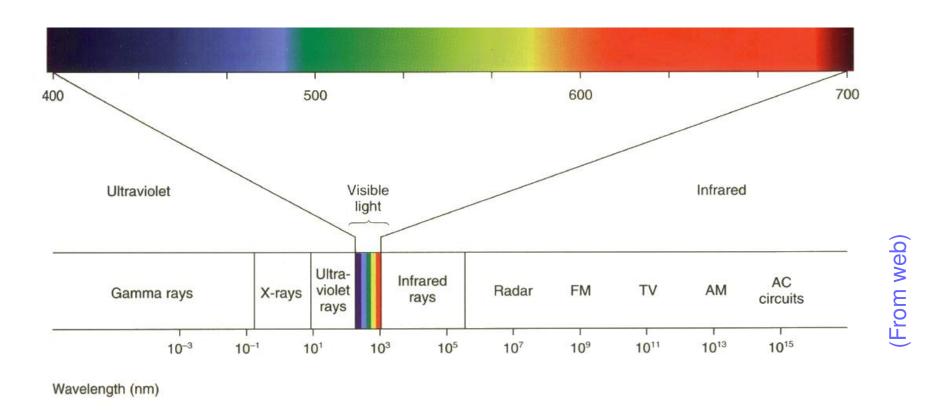
**Physiology** What is the behavior of the component parts of the visual system?

**Electrophysiology** What is the electrical behavior of neurons, measured with an electrode?

**Imaging** What is the behavior of a large area of the nervous system?

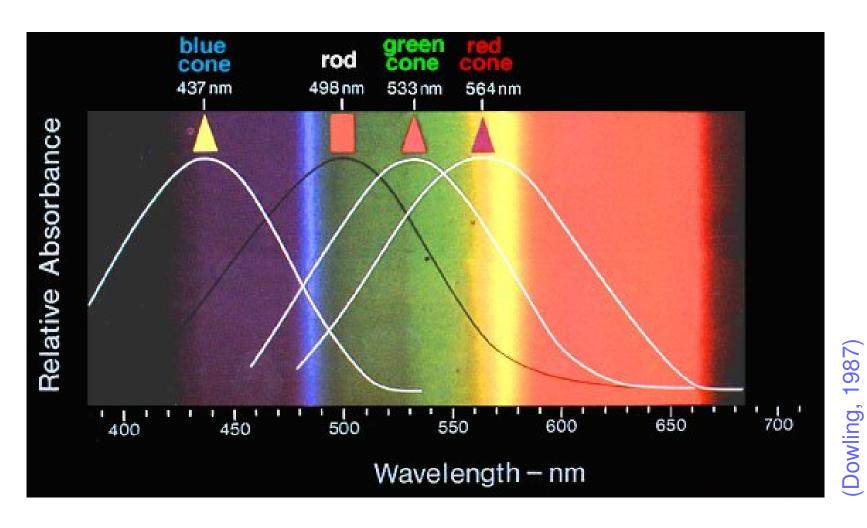
**Genetics** Which genes control visual system development and function, and what do they do?

#### Electromagnetic spectrum



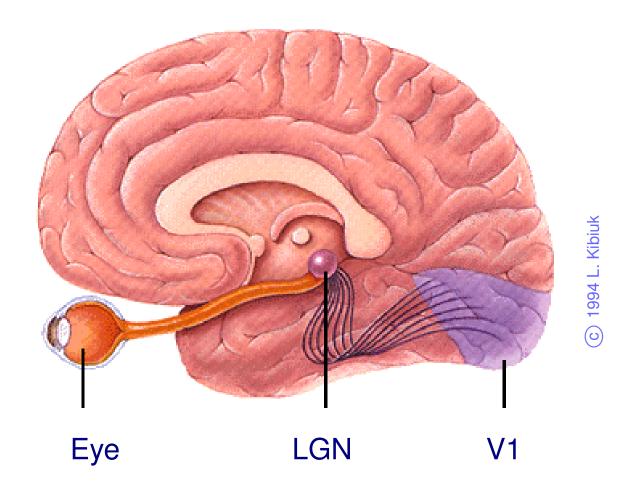
Start with the physics: visible portion is small, but provides much information about biologically relevant stimuli

#### Cone spectral sensitivities



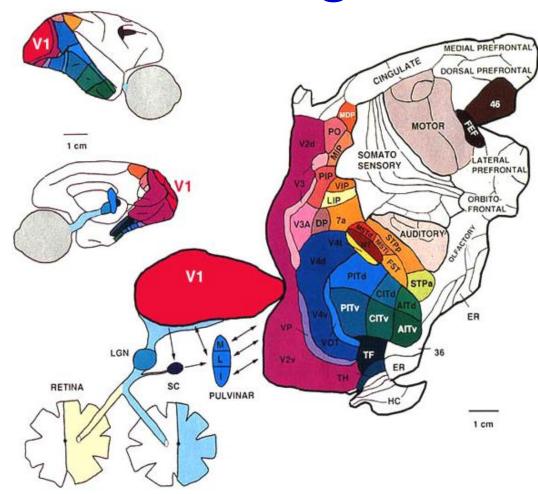
Somehow we make do with sampling the visible range of wavelengths at only three points (3 cone types)

# **Early visual pathways**



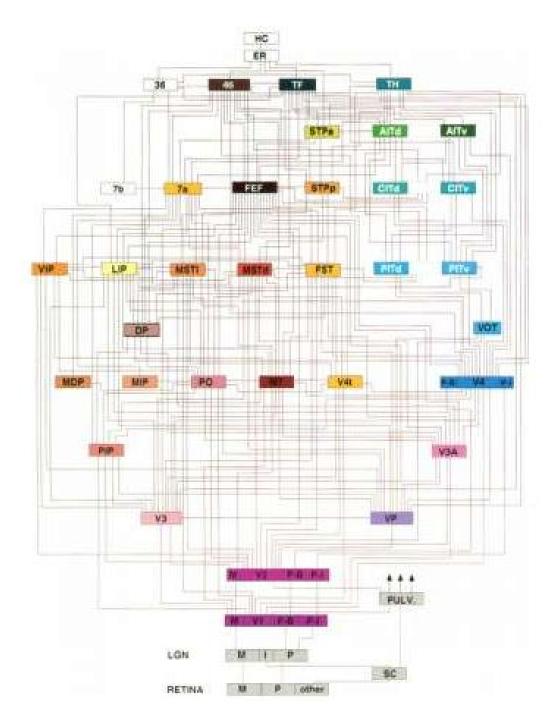
Signals travel from retina, to LGN, then to primary visual cortex

#### **Higher areas**



Macaque monkey visual areas (Van Essen et al. 1992)

- Many higher areas beyondV1
- Selective for faces, motion, etc.
- Not as well understood



# Circuit diagram

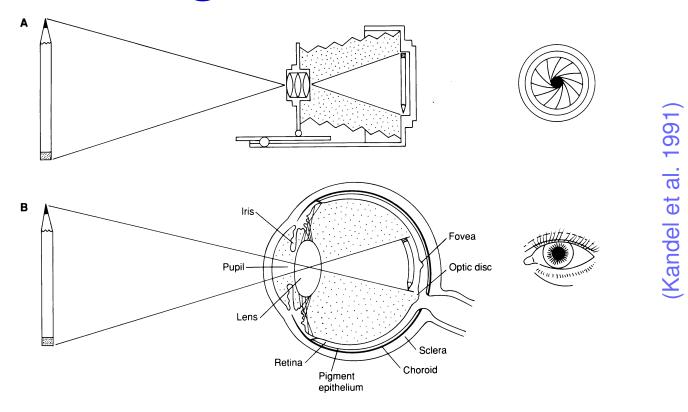
Connections between macaque monkey visual areas

(Van Essen et al. 1992)

A bit messy!

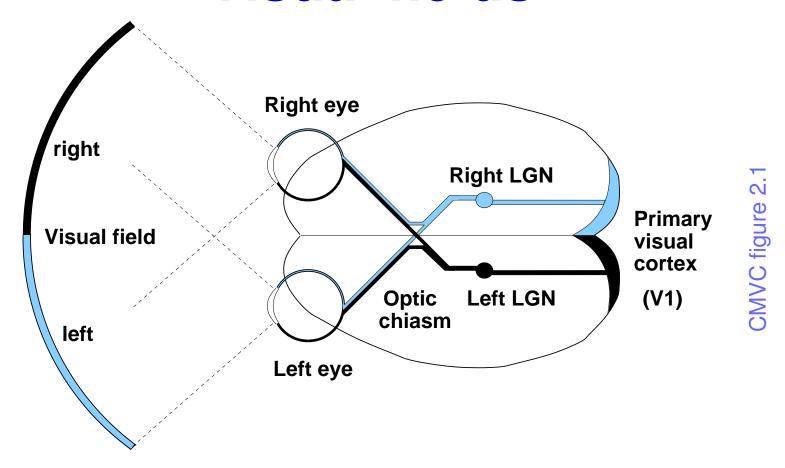
(Yet still just a start.)

# **Image formation**



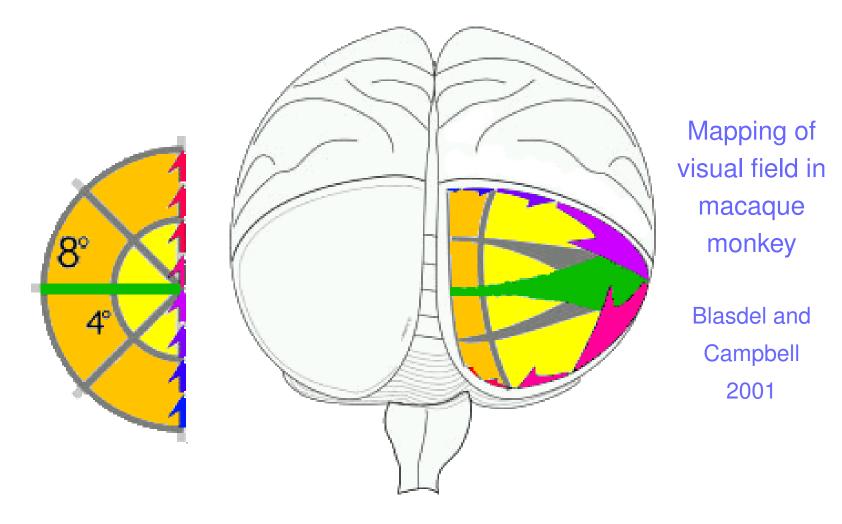
	Fixed	Adjustable	Sampling
Camera:	lens shape	focal length	uniform
Eye:	focal length	lens shape	higher at fovea

#### Visual fields



- Each eye sees partially overlapping areas
- Inputs from opposite hemifield cross over at chiasm

#### Retinotopic map



- Visual field is mapped onto cortical surface
- Fovea is overrepresented

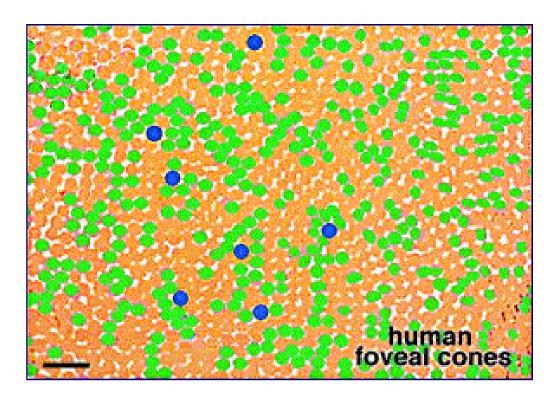
#### **Effect of foveation**

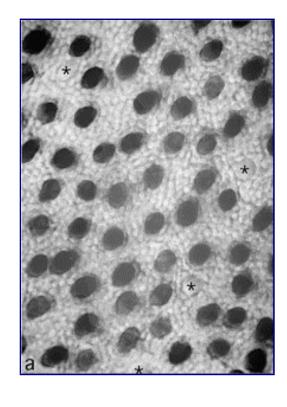




Smaller, tightly packed cones in the fovea give much higher resolution

#### Retinal surface





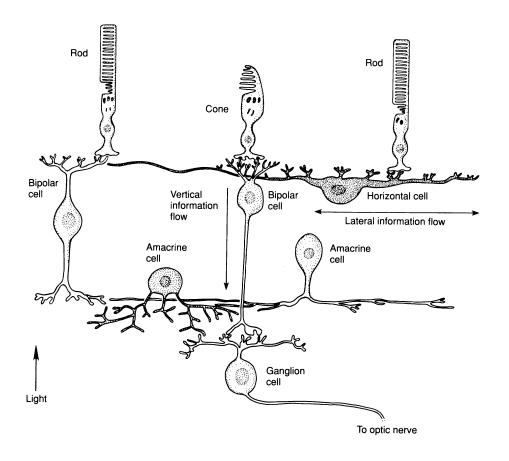
(Ahnelt & Kolb 2000); no scale in origina

Fovea (center →)

Periphery

- Fovea: densely packed L,M cones (no rods)
- No S cones in central fovea; sparse elsewhere
- Cones are larger in periphery (\*: S-cones)
- Cone spacing also increases, with gaps filled by rods

#### **Retinal circuits**

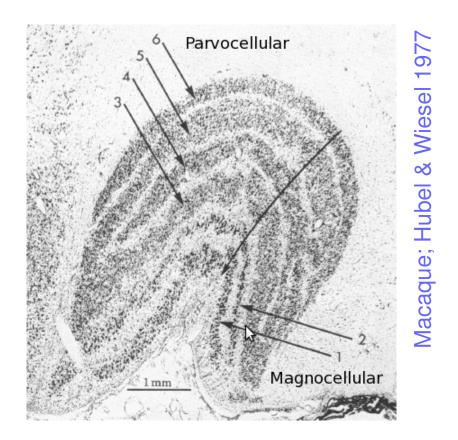


(Kandel et al. 1991)

Rod pathway Rod, rod bipolar cell, ganglion cell

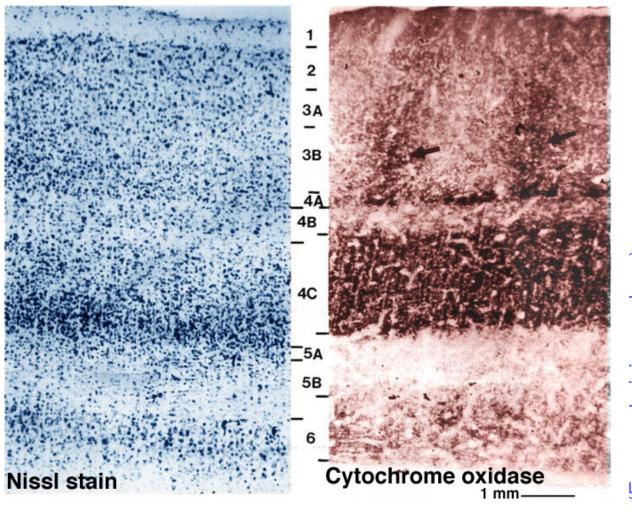
Cone pathway Cone, bipolar cell, ganglion cell

# **LGN layers**



Multiple aligned representations of visual field in the LGN for different eyes and cell types

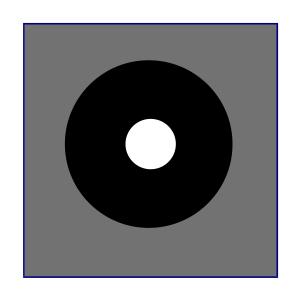
# **V1 layers**

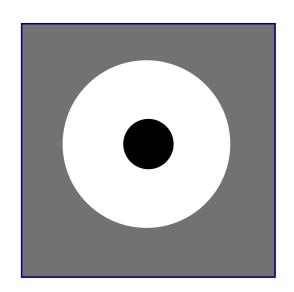


(From webvision.umh.es)

Multiple layers of cells in V1
Brodmann numbering

## Retinal/LGN cell response types





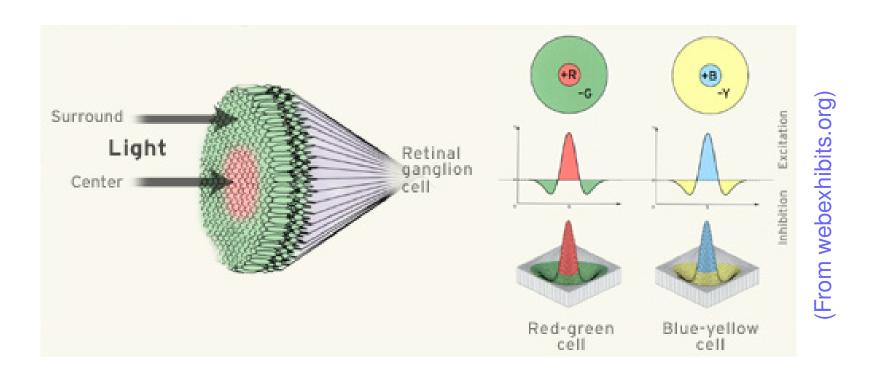
Types of receptive fields based on responses to light:

in center in surround

On-center excited inhibited

Off-center inhibited excited

#### Color-opponent retinal/LGN cells

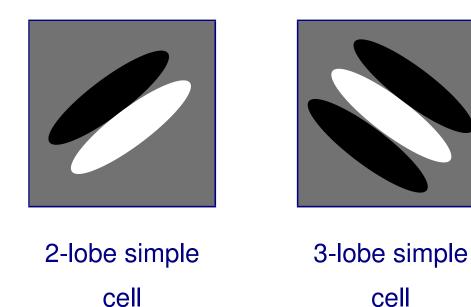


Red/Green cells: (+R,-G), (-R,+G), (+G,-R), (-G,+R)

Blue/Yellow cells: (+B,-Y); others?

Error: light arrows in the figure are backwards! Actual organization mostly consistent with random wiring

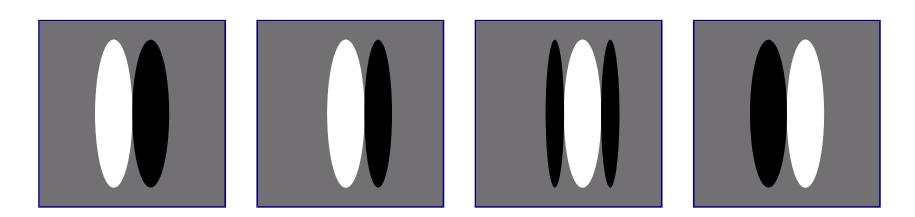
#### V1 simple cell responses



Starting in V1, only oriented patterns will cause any significant response

Simple cells: pattern preferences can be plotted as above

#### V1 complex cell responses

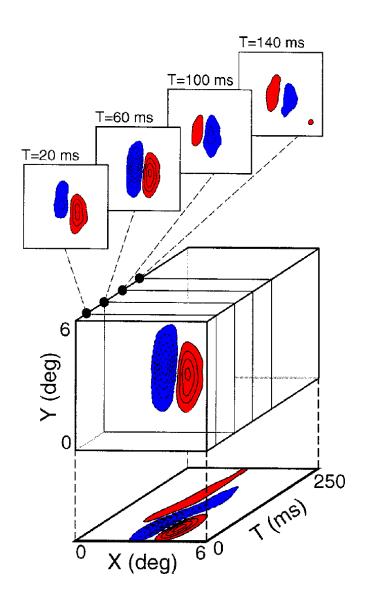


(Same response to all these patterns)

Complex cells are also orientation selective, but have responses invariant to phase

Can't measure complex RFs using pixel-based correlations

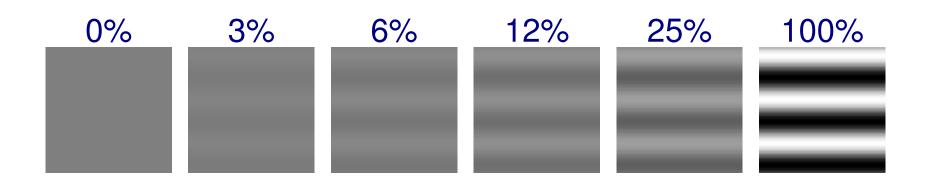
## Spatiotemporal receptive fields



- Neurons are selective for multiple stimulus dimensions at once
- Typically prefer lines moving in direction perpendicular to orientation preference

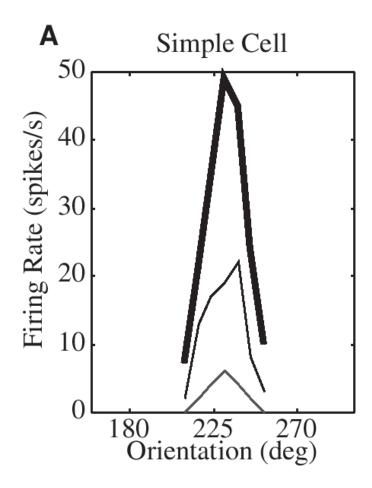
(Cat V1; DeAngelis et al. 1999)

#### **Contrast perception**



- Humans can detect patterns over a huge contrast range
- In the laboratory, increasing contrast above a fairly low value does not aid detection
- See 2AFC (two-alternative forced-choice) test in google and ROC (Receiver Operating Characteristic) in Wikipedia for more info on how such tests work

#### **Contrast-invariant tuning**



(Sclar & Freeman 1982)

- Single-cell tuning curves are typically Gaussian
- 5%, 20%, 80% contrasts
   shown
- Peak response increases, but
- Tuning width changes little
- Contrast where peak is reached varies by cell

#### **Definitions of contrast**

Luminance (luminosity): Physical amount of light

Contrast: Luminance relative to background levels to which the visual system has become adapted

Contrast is a fuzzy concept – clear only in special cases:

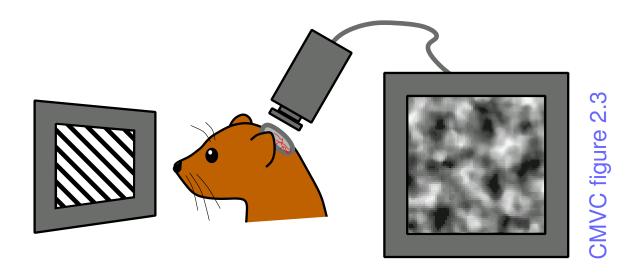
Weber contrast (e.g. a tiny spot on uniform background)

$$C = \frac{Lmax - Lmin}{Lmin}$$

Michelson contrast (e.g. a full-field sine grating):

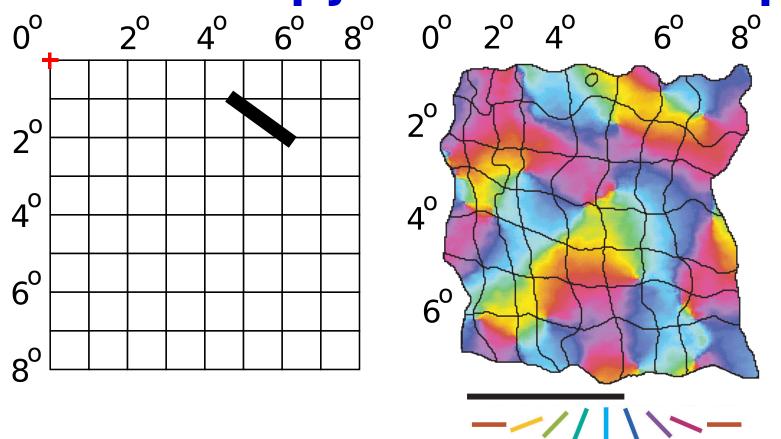
$$C = \frac{Lmax - Lmin}{Lmax + Lmin} = \frac{\frac{Lmax - Lmin}{2}}{Lavg}$$

#### Measuring cortical maps



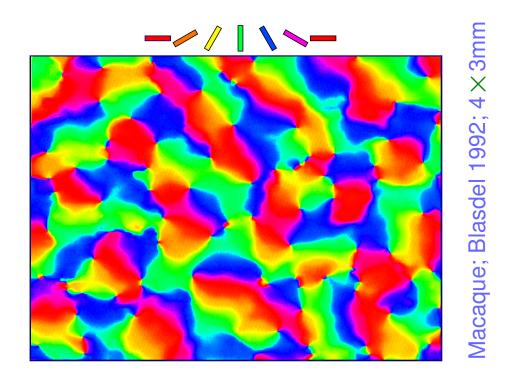
- Surface reflectance (or voltage-sensitive-dye emission) changes with activity
- Measured with optical imaging
- Preferences computed as correlation between measurement and input

#### Retinotopy/orientation map



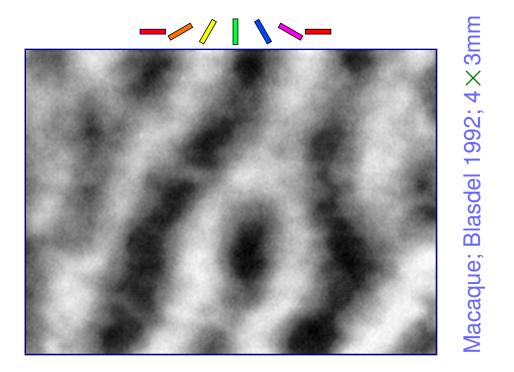
- Tree shrew has no fovea → isotropic map
- All orientations represented for each retina location
- Orientation map is smooth, with local patches

#### **Macaque orientation map**



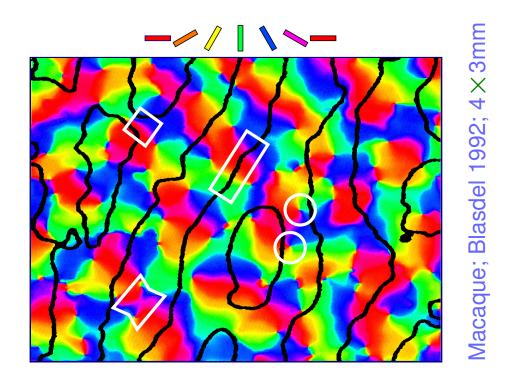
- Macaque monkey has fovea but similar orientation map
- Retinotopic map (not measured) highly nonlinear

#### Ocular dominance map in V1



- Most neurons are binocular, but prefer one eye
- Eye preference alternates in stripes or patches

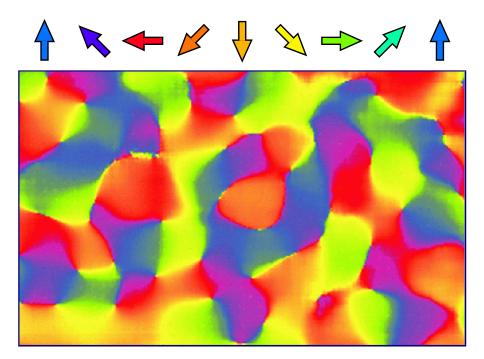
#### Combined OR/OD map in V1

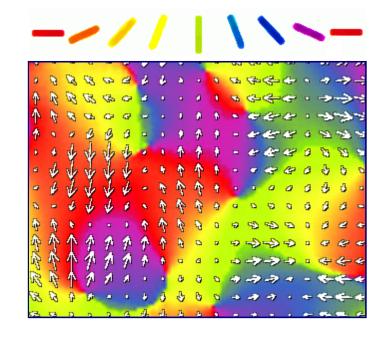


- Same neurons have preference for both features
- OR has linear zones, fractures, pinwheels, saddles
- OD boundaries typically align with linear zones

# (Adult ferret; Weliky et al. 1996)

#### **Direction map in V1**





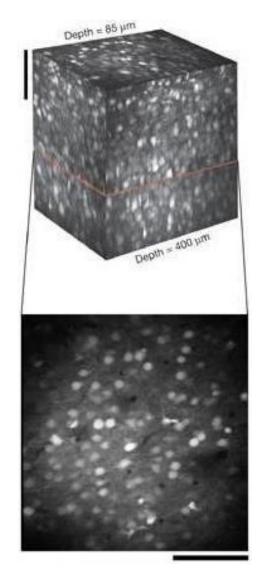
Direction preference

OR/Direction pref.

 $(3.2 \times 2mm)$ 

 $(1 \times 1.4 \text{mm})$ 

- Local patches prefer different directions
- Single-OR patches often subdivided by direction
- Other maps: spatial frequency, color, disparity

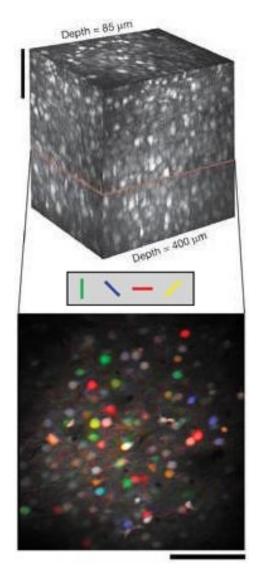


Rat V1 (scale bars 0.1mm)

#### Two-photon microscopy:

- New technique with cell-level resolution
- Can measure a small volume very precisely

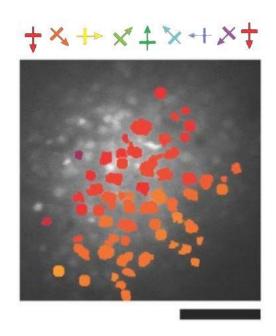
(Ohki et al. 2005)

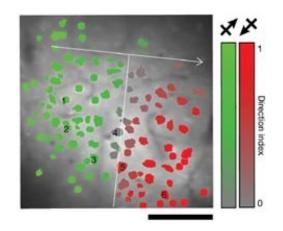


Rat V1 (scale bars 0.1mm)

- Individual cells can be tagged with feature preference
- In rat, orientation preferences are random
- Random also expected in mouse, squirrel

(Ohki et al. 2005)

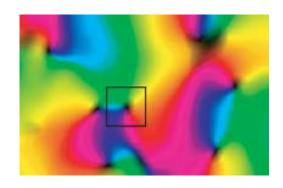




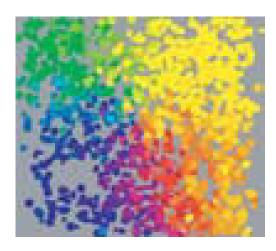
Cat V1 Dir. (scale bars 0.1mm)

- In cat, validates results from optical imaging
- Smooth organization for direction overall
- Sharp, well-segregated discontinuities

(Ohki et al. 2005)



Low-res map (2×1.2mm)

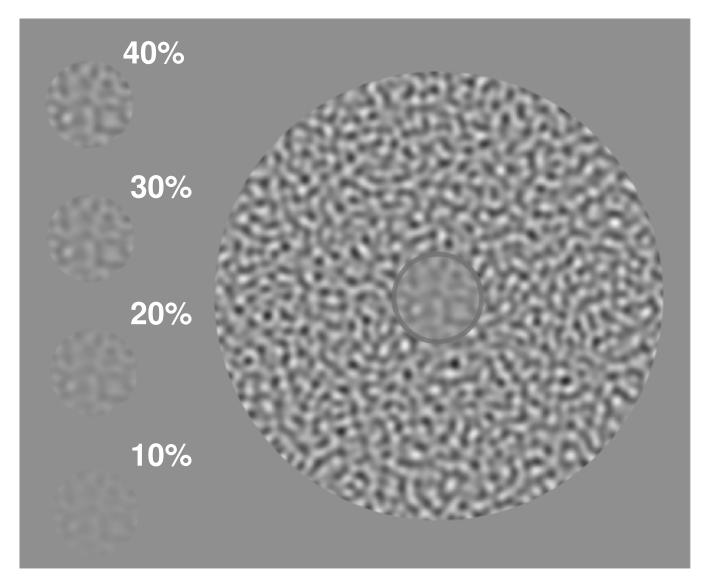


Stack of all labeled cells (0.6 × 0.4mm)

- Very close match with optical imaging results
- Stacking labeled cells from all layers shows very strong ordering spatially and in across layers
- Selectivity in pinwheels controversial; apparently lower

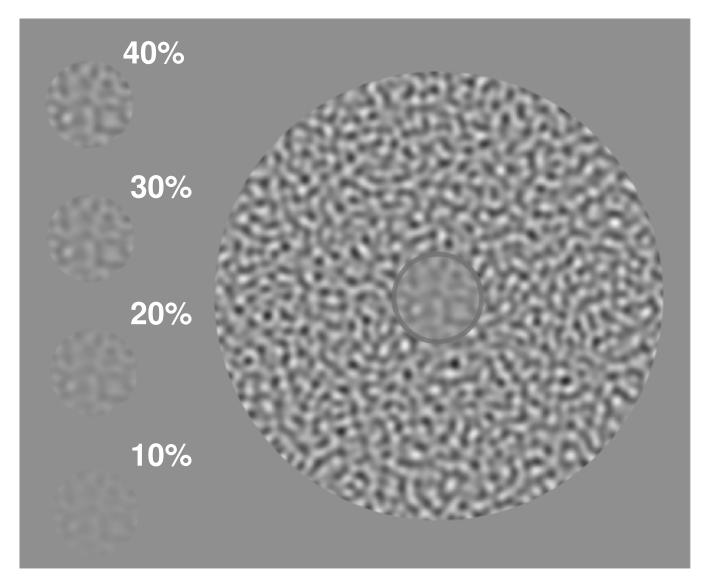
(Ohki et al. 2006)

#### **Surround modulation**



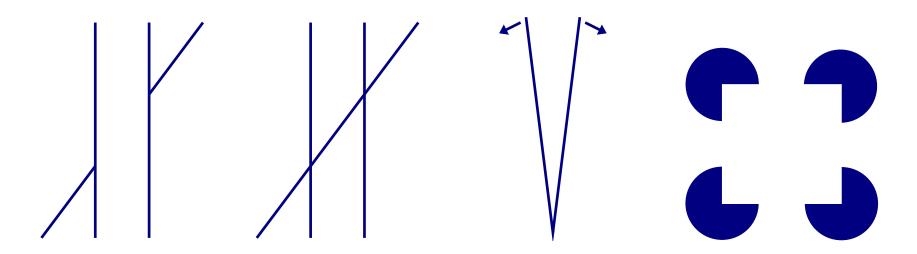
Which of the contrasts at left matches the central area?

#### **Surround modulation**



Which of the contrasts at left matches the central area? 40%

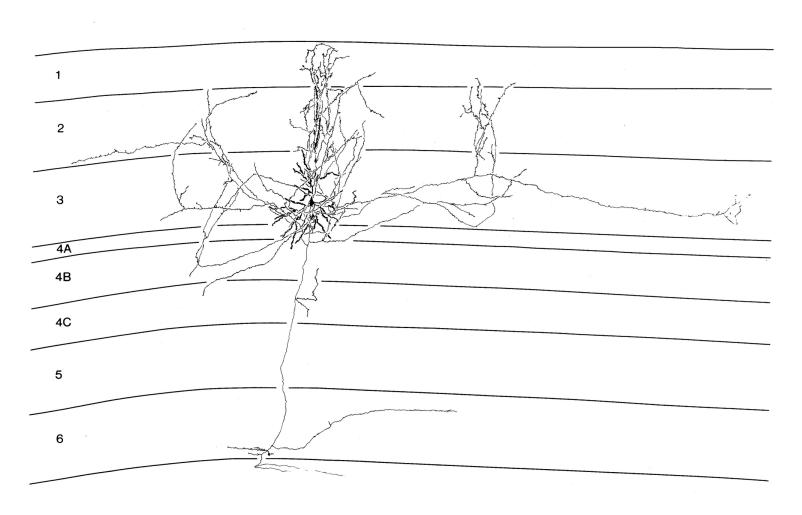
### **Contextual interactions**



- Orientation and shape perception is not entirely local (e.g. due to individual V1 neurons).
- Instead, adjacent line elements interact (tilt illusion).
- Presumably due to lateral or feedback connections at V1 or above.

# (Macaque; Gilbert et al. 1990)

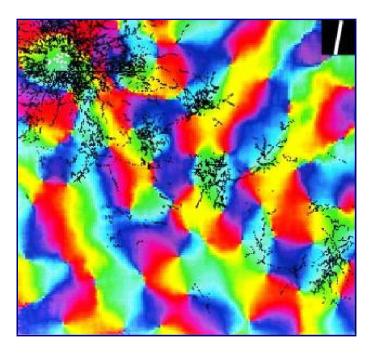
### **Lateral connections**



- Example layer 2/3 pyramidal cell
- Patchy every 1mm

### Lateral connections

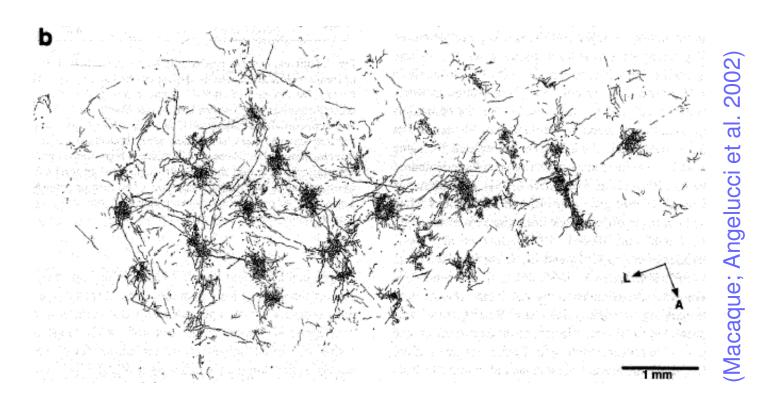




(2.5 mm × 2 mm in tree shrew V1; Bosking et al. 1997)

- Connections up to 8mm link to similar preferences
- Patchy structure, extend along OR preference

### Feedback connections



- Relatively little known about feedback connections
- Large number, wide spread
- Some appear to be diffuse
- Some are patchy and orientation-specific

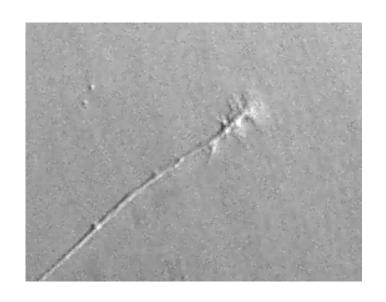
# Visual development

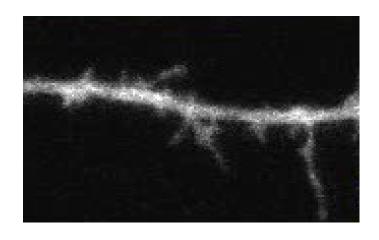
Research questions studied in this course:

- Where does the visual system structure come from?
- How much of the architecture is specific to vision?
- What influence does the environment have?
- How plastic is the system in the adult?

Most visual development studies focus on ferrets and cats, whose visual systems are very immature at birth.

# **Initial development**





(Ziv 1996

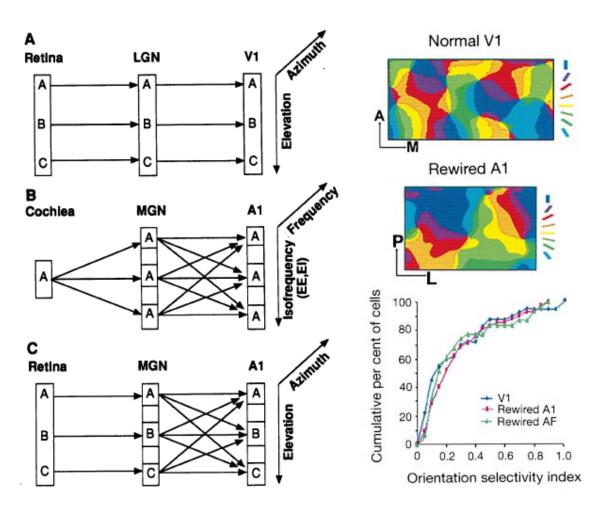
- Tissues develop into eye, brain
- RGC axons grow from eye to LGN and superior colliculus (SC) following chemical gradients
- Axons form synapses at LGN, SC
- LGN axons grow to V1, V2, etc., forming synapses

# **Cortical development**

- Coarse cortical architecture (e.g. division into areas)
   appears to be fixed after birth
- Cortical architecture similar across areas
- Much of cortical development appears driven by different peripheral circuitry (auditory, visual, etc.)
- E.g. Sur et al. (1998-2000): auditory cortex can develop into visual cortex

### **Rewired ferrets**

### Sur et al. 1988-2000:



- Disrupt
   connections
   to MGN
- RGC axons now terminate in MGN
- 3. Then to A1 instead of V1
- 4. → Functionalorientation cells,map in A1

# Human visual system at birth

- Some visual ability
- Fovea barely there
- Color vision poor
- Binocular vision difficult
  - Poor control of eye movements
  - Seems to develop later
- Acuity increases 25X (birth to 6 months)

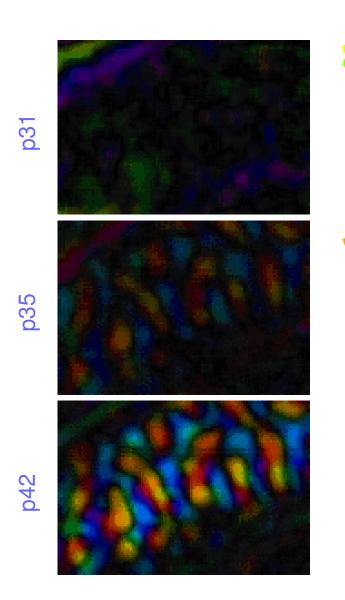
# Map development

- Initial orientation, OD maps develop without visual experience (Crair et al. 1998)
- Maps match between the eyes even without shared visual experience (Kim & Bonhoeffer 1994)
- Experience leads to more selective neurons and maps (Crair et al. 1998)
- Lid suture (leaving light through eyelids) during critical period destroys maps (White et al. 2001)
- → Complicated interaction between system and environment.

# **OR** map development

Ferret; Chapman et al. 1996)

 $5mm \times 3.5mm$ 

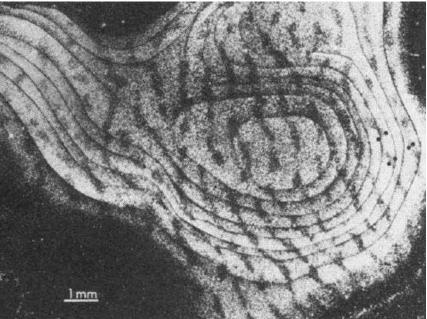


Map not visible when
 eyes first forced open

Cradually becomes

- Gradually becomes stronger over weeks
- Shape doesn't change significantly
- Initial development
   affected little by dark
   rearing

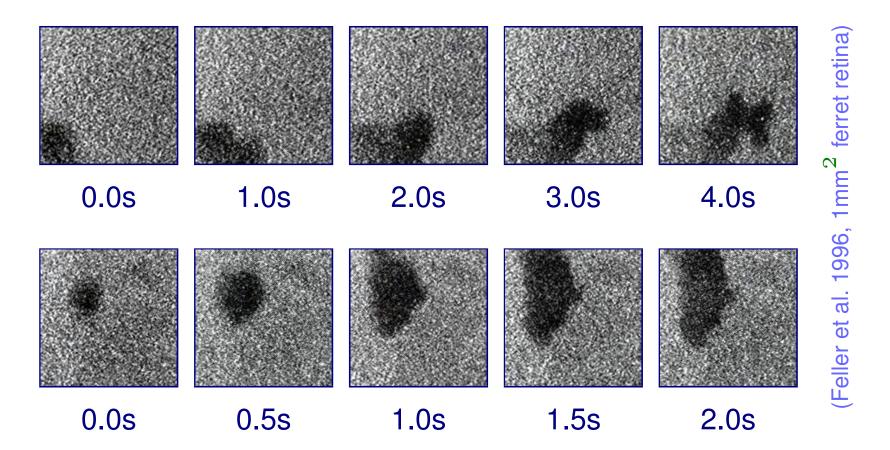
# Monocular deprivation



- (Left eye (open) labeled white)
- Raising with one eyelid sutured shut results in larger area for other eye
- Sengpiel et al.
   1999: Area for overrepresented orientations increases too

(Monkey V1 layer 4C; Wiesel 1982)

# Internally generated inputs

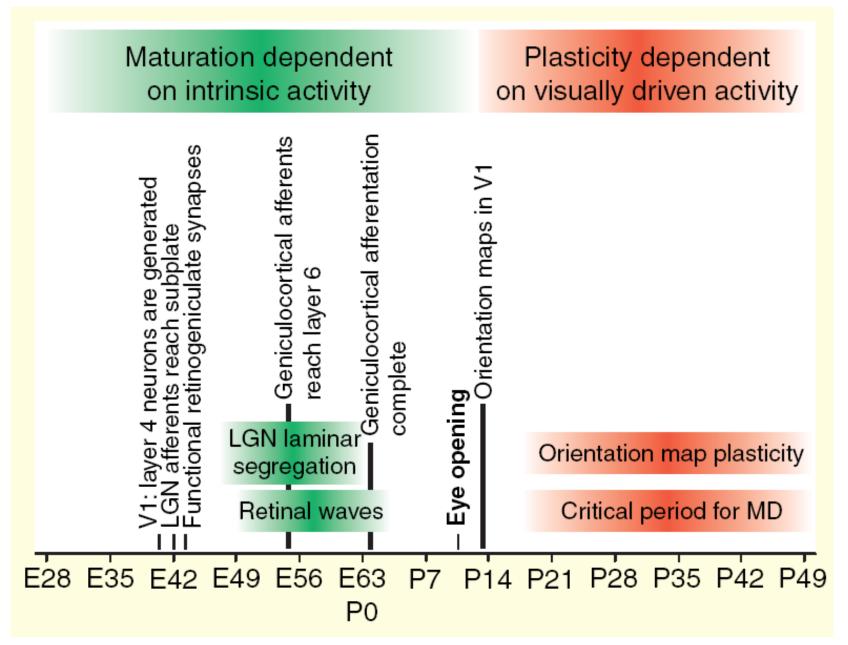


- Retinal waves: drifting patches of spontaneous activity
- Training patterns?

# Role of spontaneous activity

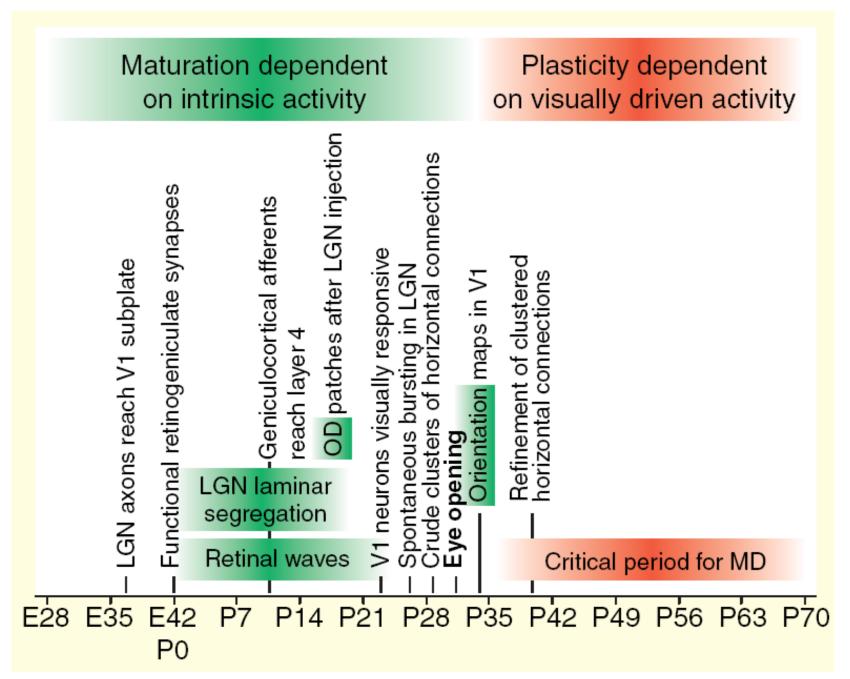
- Silencing of retinal waves prevents eye-specific segregation in LGN (Huberman et al. 2003) and ocular dominance columns in V1 (Huberman et al. 2006)
- Boosting in one eye disrupts LGN, but not if in both
- Disrupting retinal waves disrupts geniculocortical mapping (Cang et al. 2005)
- Other sources of input to V1: spontaneous cortical activity, brainstem activity
- All developing areas seem to be spontaneously active,
   e.g. auditory system, spinal cord

### **Timeline: Cat**

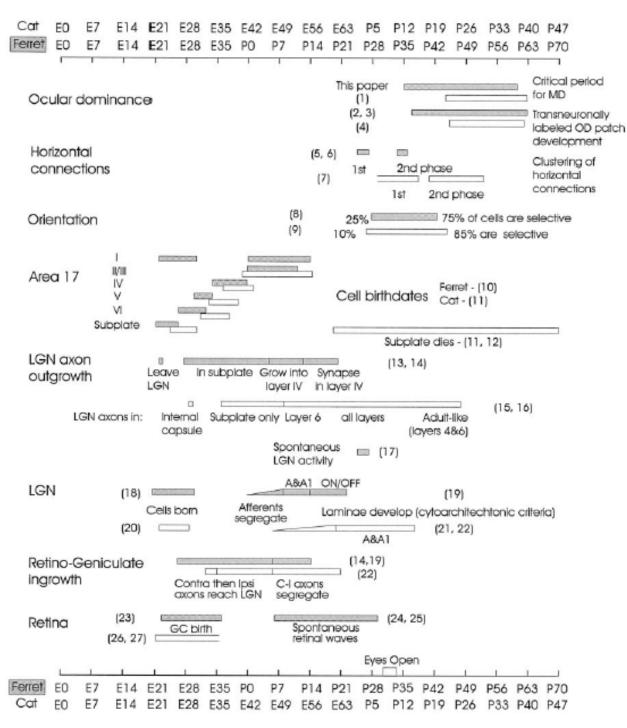


(Sengpiel & Kind 2002)

# **Timeline: Ferret**



(Sengpiel & Kind 2002)



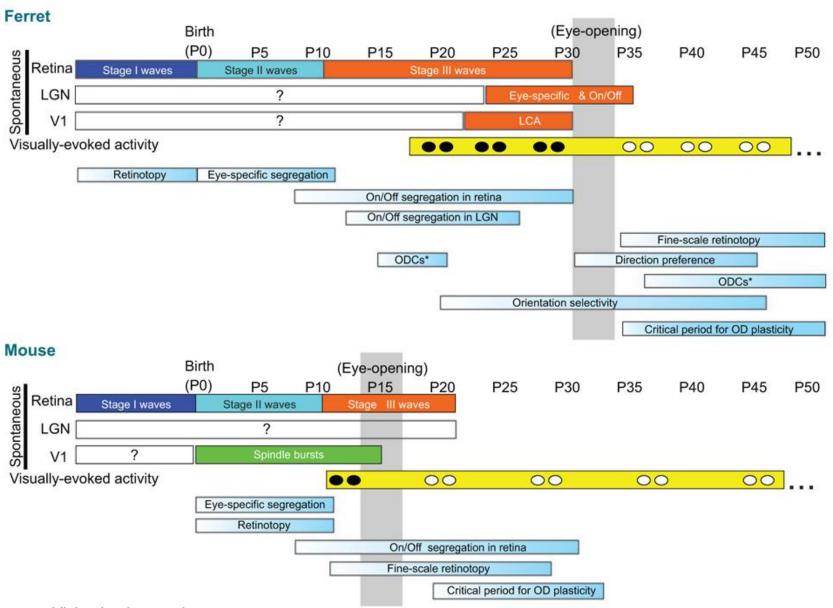
# Cat vs. ferret

Should be readable in a printout, not on screen

OD, Ocular dominance
MD, monocular deprivation
GC, ganglion cell
C-I, contralateral-ipsilateral

(Issa et al. 1999)

### Ferret vs. mouse



(Huberman et al. 2008)

## **Conclusions**

- Early areas well studied
- Higher areas much less so
- Little understanding of how entire system works together
- Development also a mystery
- Lots of work to do

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