Exploring Variation in Biochemical Pathways with the Continuous π -Calculus

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The continuous π -calculus ($c\pi$) is a process algebra for modelling behaviour and variation in biomolecular systems: *e.g.* enzyme activation and inhibition; circadian clocks; signalling pathways.

Expressions in $c\pi$ represent mixtures of chemical reagents, and can be compiled to conventional ODE models for fast numerical simulation.

With a language of potential changes in $c\pi$ processes we systematically explore evolutionary neighbourhoods of a specific signalling pathway, and observe instances of robustness, neutrality and evolvability.

A complementary temporal logic for behaviour in context gives a language to classify these variations in behaviour.



Marek Kwiatkowski and Ian Stark.

On Executable Models of Molecular Evolution. In *Proc. 8th International Workshop on Computational Systems Biology WCSB 2011*, pp. 105–108.

Continuous π is a name-passing process algebra for modelling behaviour and variation in molecular systems.

Based on Milner's π -calculus, it introduces real-valued variability in:

- rates of reaction;
- affinity between interacting names; and
- quantities of processes.

Although sharing an approach common to process algebras for biomodelling, some features are distinctive. For example, by comparison with the stochastic pi-calculus:

- ODEs are the primary mode of execution, not stochastic simulation
- Continuous concentrations of chemicals replace discrete individuals
- End-to-end channels are replaced by multiple competing names

Continuous π has two levels of system description:

- Species
 - Individual molecules (proteins)
 - Transition system semantics
- Processes
 - Bulk population (concentration)
 - Differential equations

Process space arises as a real-valued vector space over species, with each point the state of a system and behaviours as trajectories through that.

For individual species, continuous π uses a modelling idiom based on that of Reggie and Shapiro:

- Reagent-centric rather than rule-based
- Individual species are represented by processes
- Complexes are modelled by name restriction vx.(A | B)
- Interaction is modelled by communication between names
- ...but with competition between multiple alternatives

As in standard π -calculus, *names* indicate a potential for interaction: for example, the docking sites on an enzyme where other molecules may attach.

These sites may interact with many different other sites, to different degrees.

This variation is captured by an *affinity network*: a graph setting out the interaction potential between different names.



Kuhn et al. Cell (2007:1260-72)

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Take a language for interaction between individual species and raise it into one for reactions in mixtures:

We can identify processes with elements of *process space* $\mathcal{P} = \mathbb{R}^{S}$, where S is the set of species.

Process Semantics

- $\frac{dP}{dt}$: Immediate behaviour
 - $\bullet~$ Vector field $\frac{d}{dt}$ over process space ${\cal P}$
 - Equivalent to an ODE system

- ∂P : Interaction potential
 - Captures available reactivity

• Element of
$$\mathbb{R}^{\mathcal{N} \times \mathcal{S} \times \mathcal{C}}$$

$$\partial(P \parallel Q) = \partial P + \partial Q$$
$$\frac{d(P \parallel Q)}{dt} = \frac{dP}{dt} + \frac{dQ}{dt} + \partial P \oplus \partial Q$$

Both $\frac{dP}{dt}$ and ∂P are defined by induction on the structure of processes; and beneath that, from the transitions of component species $c \cdot A$.

With this, we are able to compose the phase portraits of our systems.

Example: Enzyme Catalysis



$$\begin{split} S &= s(x, y).(x.S + y.(P|P')) \\ E &= \nu(u, r, t: M).(e\langle u, r \rangle.t.E) \\ P &= P' = \tau @k_{degrade}.0 \\ c_S \cdot S \parallel c_E \cdot E \end{split}$$



Process Space: Substrate & Product & Enzyme



Example: Synechococcus Elongatus



S. Elongatus circadian clock proteins, effective in vitro: KaiA, KaiB and KaiC. (Tomita et al. 2005)

Several mechanisms have been proposed: one is the cyclic six-fold phosphorylation of KaiC hexamers in two alternative conformations, stabilised by KaiA and KaiB. (van Zon et al. 2007)





Execution and Modification



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Process Algebras for Molecular Evolution

One way to model molecular evolution is by specific modifications of concrete mathematical models.

Process algebras, and similar intermediate languages, offer a framework to generalise this model for variation and selection.

Process ~ Genotype Execution ~ Development Behaviour ~ Phenotype

Relevant features of models like continuous π include:

- Reagent-centric models to match genetic variation
- Free formation of new terms, particularly novel complexes
- Computable behaviour of created components

Variation operators are transformations of $c\pi$ models which correspond to evolutionary events.

Ideally, a suite of such operations should:

- Maintain the biological idiom
- Be biologically meaningful
- Be expressive enough to build new reaction networks from scratch

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For example: site reconfiguration



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We have defined a dozen such operators modelling gene duplications, gene knockouts, changes in activity rates within complexes, and more.

Simplified MAPK Cascade



MAPK in $c\pi$

. . .

$$Ras = (\mathbf{v}x \frown \overline{x})ras(x; y).(\overline{x}.Ras + y.Ras)$$
$$Raf = (\mathbf{v}x \frown \overline{x})raf(x; y).(\overline{x}.Raf + y.Raf^*)$$

$$ERK^{**} = (\mathbf{v}\mathbf{x} \frown \overline{\mathbf{x}})erk_{\mathbf{b}}^{**}(\mathbf{x};\mathbf{y}).(\overline{\mathbf{x}}.ERK^{**} + \mathbf{y}.ERK^{*})$$
$$MKP3 = (\mathbf{v}\mathbf{x} \frown \overline{\mathbf{x}})mkp3(\mathbf{x};\mathbf{y}).(\overline{\mathbf{x}}.MKP3 + \mathbf{y}.MKP3)$$

$$\Pi = c_1 \cdot \textit{Raf} \parallel c_2 \cdot \textit{Ras} \parallel \ldots \parallel c_4 \cdot \textit{ERK} \parallel c_7 \cdot \textit{MKP3}$$



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MAPK Behaviour



This MAPK model compiles into 23 differential equations, which are then solved with Octave. The signalling cascade correctly transmits initial presence of **Ras** into a peak of **ERK**** via **Raf*** and **MEK****.

Evolutionary Analysis of MAPK



- Reconfigure every site in every way possible $(16 \times 2^{16} \approx 10^6)$.
- Generate ODEs and hence behaviour traces for every variant.

Qualitative analysis

- Classify phenotypes with LTL model-checking
- Find evolutionarily fragile and robust sites

Quantitative analysis

- Compute the fitness of every variant using signal integration
- Find the distribution of mutation effects on fitness

Phenotype classes

- Four categories: peak, switch, oscillatory, noise.
- Automatically identified using LTL checking.
- Results: peak 7.0%; switch 45.2%; oscillatory 0.0; noise 47.8%.

Fitness



Fitness is the area marked green minus the area marked red.

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Fitness Distribution



Histogram with 500 evenly-sized bins; green sections are *peak* variants; red vertical line shows initial model.

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Fitness Distributions by Site Modified



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Less Fit Peaks (Left)



Less Fit Peaks (Right)



Advantageous Mutations



We have been able to explore the complete one-step evolutionary neighbourhood of a MAPK cascade under modifications of site activity.

For this model, we observe:

- Signal transmission has some robustness.
- Switch behaviour is readily accessible.
- Almost all mutations reduce fitness, although many only slightly so.
- A few give improvement against the chosen fitness measure.

Logic for Behaviour in Context

To complement systematic operations for variation in processes, we give a language for classifying the resulting behaviours:

 $P \models b$

Process P exhibits behaviour b

Our language is a temporal logic with real-valued constraints and behaviour in context:

Basic observationsCoLogical operatorsbBehaviour over timeFiTime-limited behaviourFiBehaviour in context(Co

$$\begin{split} & \text{Concentration } [A] > c \text{, rate of change } [B]' < k \\ & b \wedge b', \ \neg b, \ \ldots \\ & \text{F}(b), \ \text{G}(b), \ b_1 \ \text{U} \ b_2, \ \ldots \\ & \text{F}_t(b), \ \ldots \\ & (Q \triangleright b) \end{split}$$

$$\mathsf{P} \models (\mathsf{Q} \triangleright \mathfrak{b}) \quad \Longleftrightarrow \quad (\mathsf{P} \parallel \mathsf{Q}) \models \mathfrak{b}$$

$$\begin{split} \mathsf{F}([Mek*] > c) \\ \mathsf{G}(([Raf] > 200) \lor ([Raf*] > 200)) \\ \mathsf{G}(\mathsf{F}([KaiC_6]') > 0.44) \end{split}$$

 $\mathsf{G}([\texttt{Prod}] < 5) \land (\texttt{En} \triangleright \mathsf{F}_t([\texttt{Prod}] > 20))$

 $(\mathsf{En} \triangleright \mathsf{F}_t([\mathsf{Prod}] > 20)) \land (\mathsf{Inhib} \triangleright \mathsf{G}(\neg(\mathsf{En} \triangleright \mathsf{F}([\mathsf{Prod}] > 20)))$

If En is added then within t seconds the concentration of Prod will rise above 20mM, but if instead Inhib is introduced then from that point on, addition of En will never lead to production of Prod. We can check whether process P exhibits behaviour b by:

- Compiling P to a collection of ODEs
- Solving numerically to give a trace of species concentrations over time
- Checking whether that trace satisfies b

However, this approach has limitations:

- Precision Indefinite temporal operators like G(-) and F(-) cannot always be checked with finite traces. Even for finite time operators $F_t(-)$, traces are only intermittent.
 - Cost Checking temporal operators is linear in trace length. But combining with contextual operators $G(Q \triangleright -)$ requires computation and traversal of many traces.

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Limitations

- Over-expressiveness of $c\pi$: stay within the biological
- Artificiality of behaviour modelling within complexes
- Low-count species (DNA) and discrete state transitions

Further Directions

- Explore computational cost of model-checking
- Lazier model-checking algorithms
- Other non-transcriptional clocks; bistable systems
- Hybrid models for discrete states

The Continuous π -makers



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Seeking a job in evolutionary aspects of theoretical/ computational/systems biology. Hire him, he's excellent.



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