

SBM 2

28-09-09

Bintu et al. (Hwa group)

Transcriptional regulation by the numbers: applications

next 5 classes

- Bintu 1: actual modeling and calibration of simple TF systems
 - Bintu 2: the thermodynamic model (theory of the above)
 - Elowitz*: combinatorial synthesis of Bintu-like promoters
 - model-driven design, Jim Collins
 - Marchisio & Stelling (Bioinformatics 24, 1903 - 2008):
compositional building of bio-brick models
- ... and then we do detailed modeling for another 5 classes

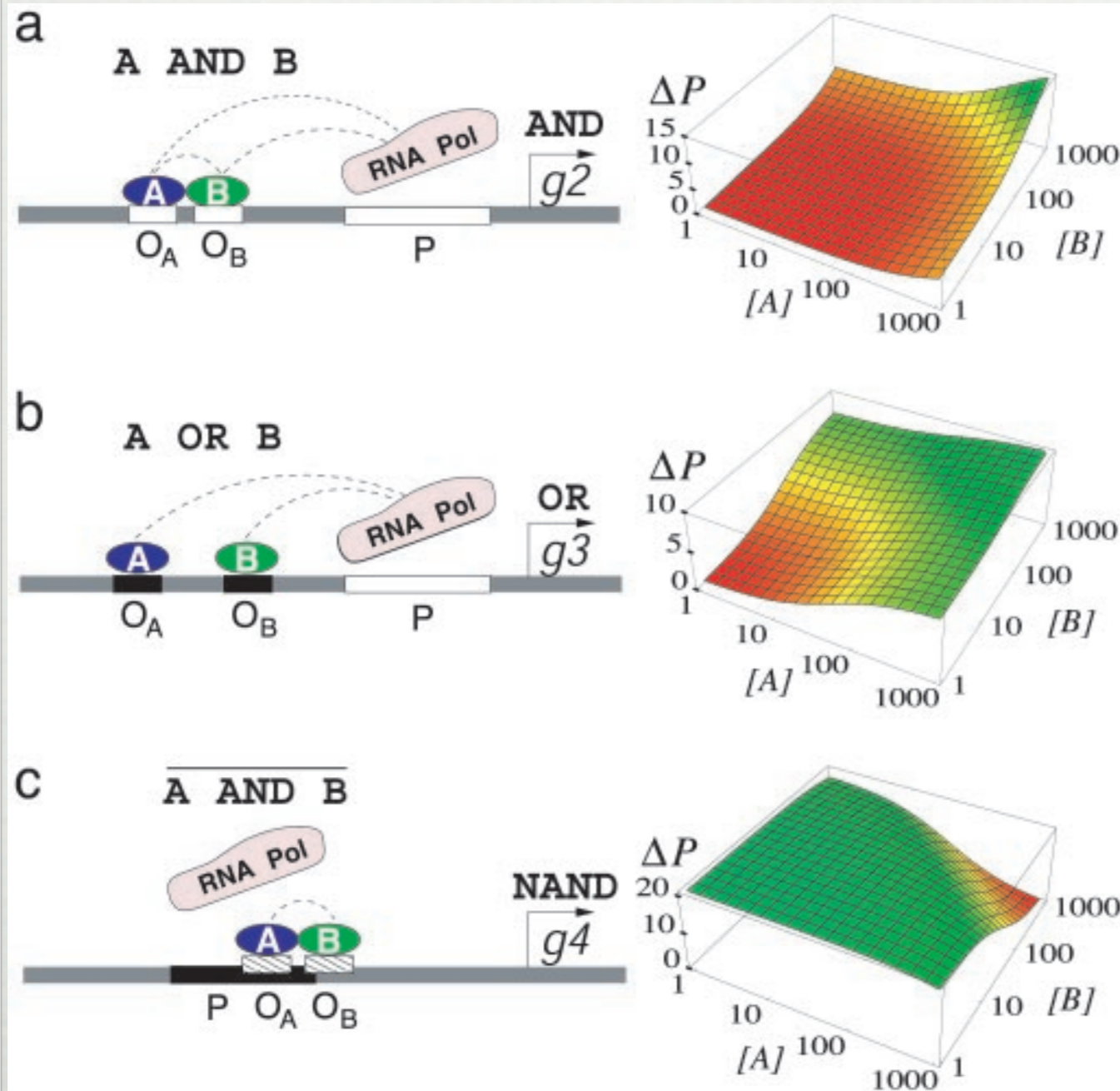
(prok) transcriptional logic - Terminology

- The activity of a gene might be regulated by that of other genes with products called transcription factors (TFs)
- depending on concentrations and other parameters TFs bind more or less regions on DNA strands upstream of a gene: called promoters
- specific TF targets on DNA are called operators (subsets of promoter, terminology varies).

(prok) transcriptional logic - Terminology 2

- TFs can pair with small molecules: CRP with cAMP, lacR with allolactose, MelR with melibiose, etc; usually this activates activators, and represses repressors (monotony)
- once in place TFs interact with the RNA polymerase (RNAP) determining the promoter activity (roughly the transcription rate, but transcription is complex in detail/more later)
- the TF active form is said to be induced, and the chemical partner is called the inducer

let us look at a few examples (theoretic) - binary "gates"

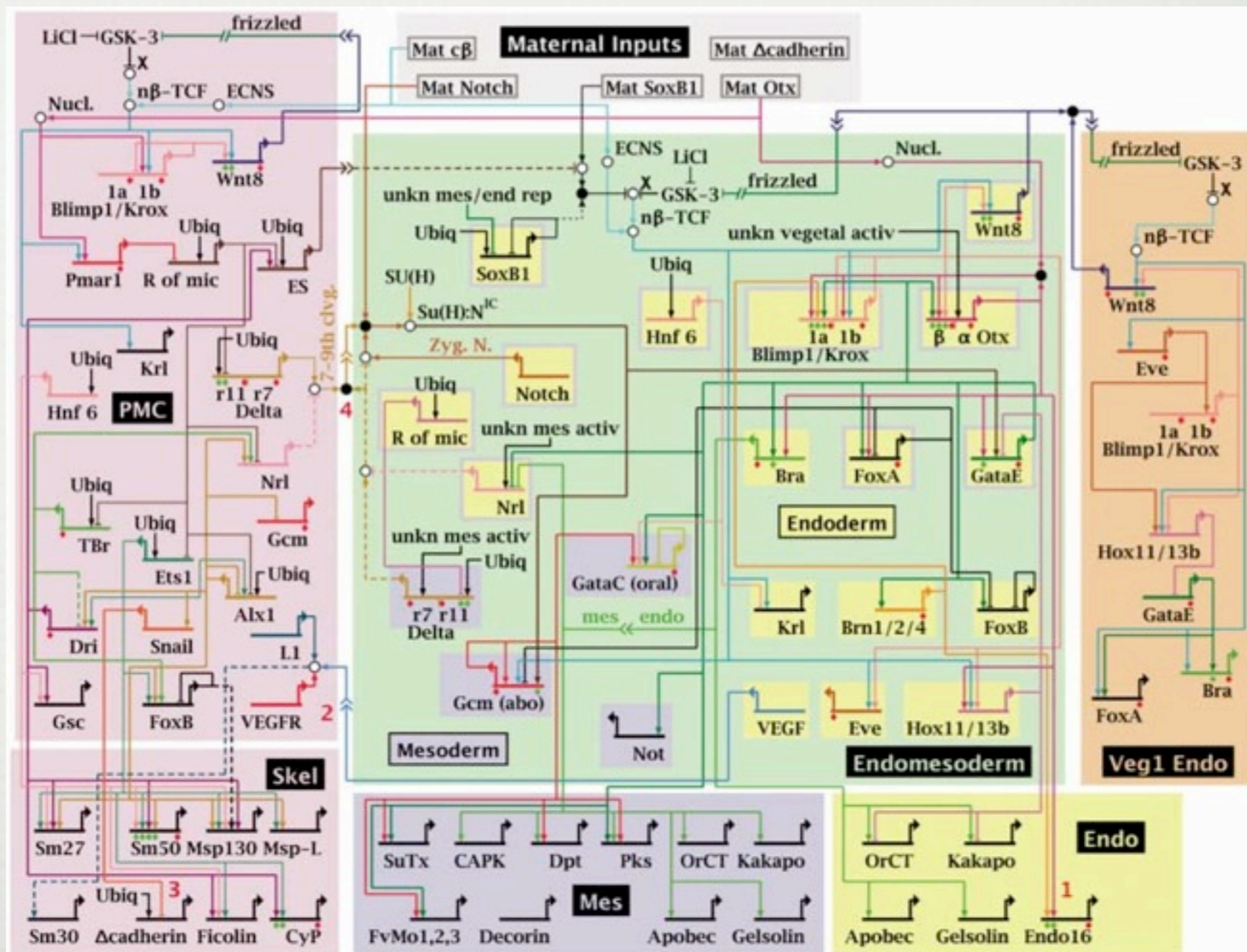


Buchler et al, PNAS
 100(9)

deciding whether
 AND or OR needs a
 discretisation -
 more later

caveat: these simple parts can be combined!

□ can become
hugely
complex in
euk.
development
(Davidson's et
al on sea
urchin)



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this class ...

various combinations of activators

- with explicit formulas for transcriptional boost, F

1. simple activation: CRP/ P_{lac}

$$F = (1 + f [A]/K_A) / (1 + [A]/K_A)$$

a ratio, ie a multiplicative factor

log-log plot:

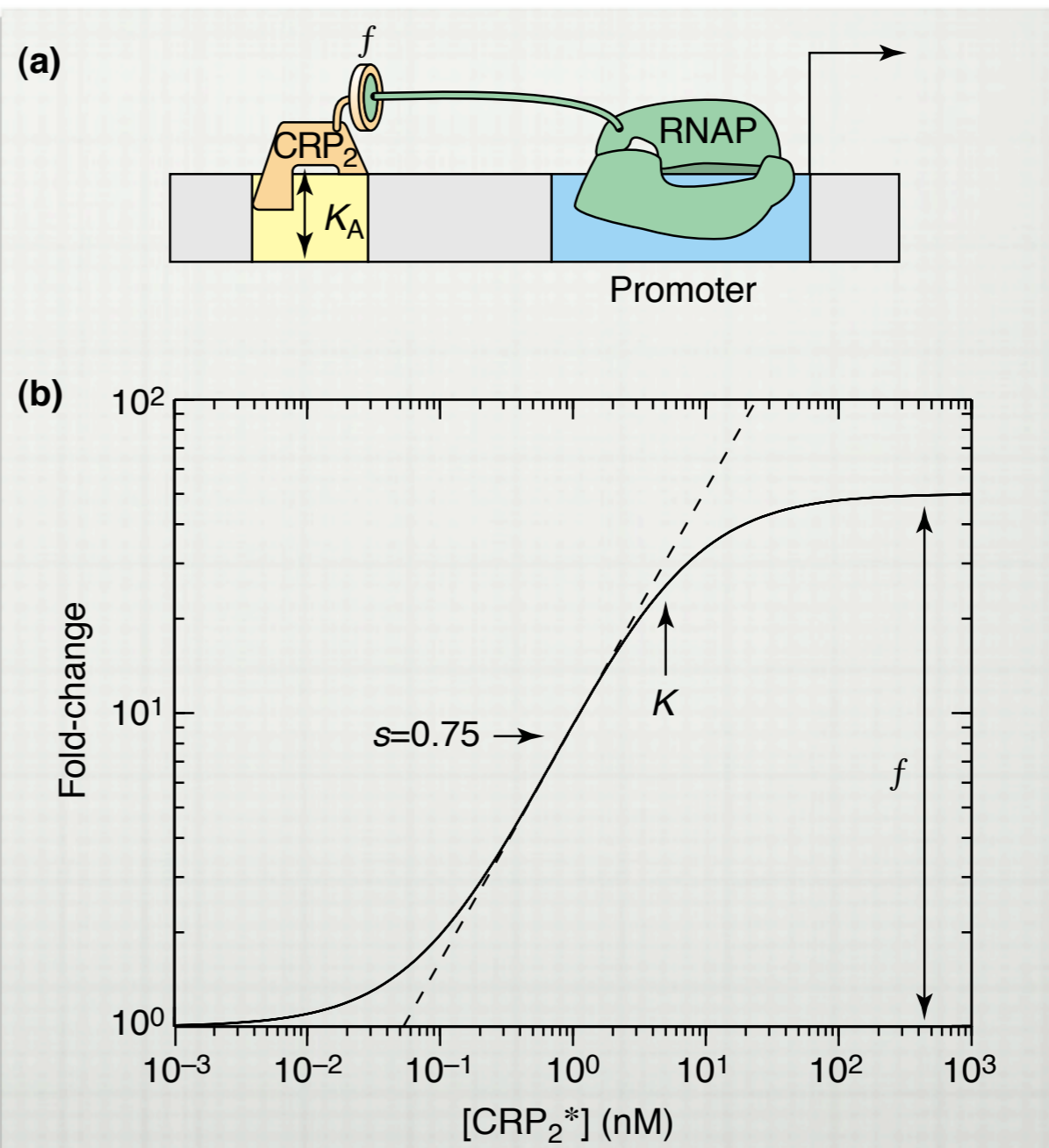
$[CRP_2^*]$ = active (dimer) TF
fold-change F

params:

K_A = eq dissociation constant

f = cooperation > 1

s = sensitivity/steepness



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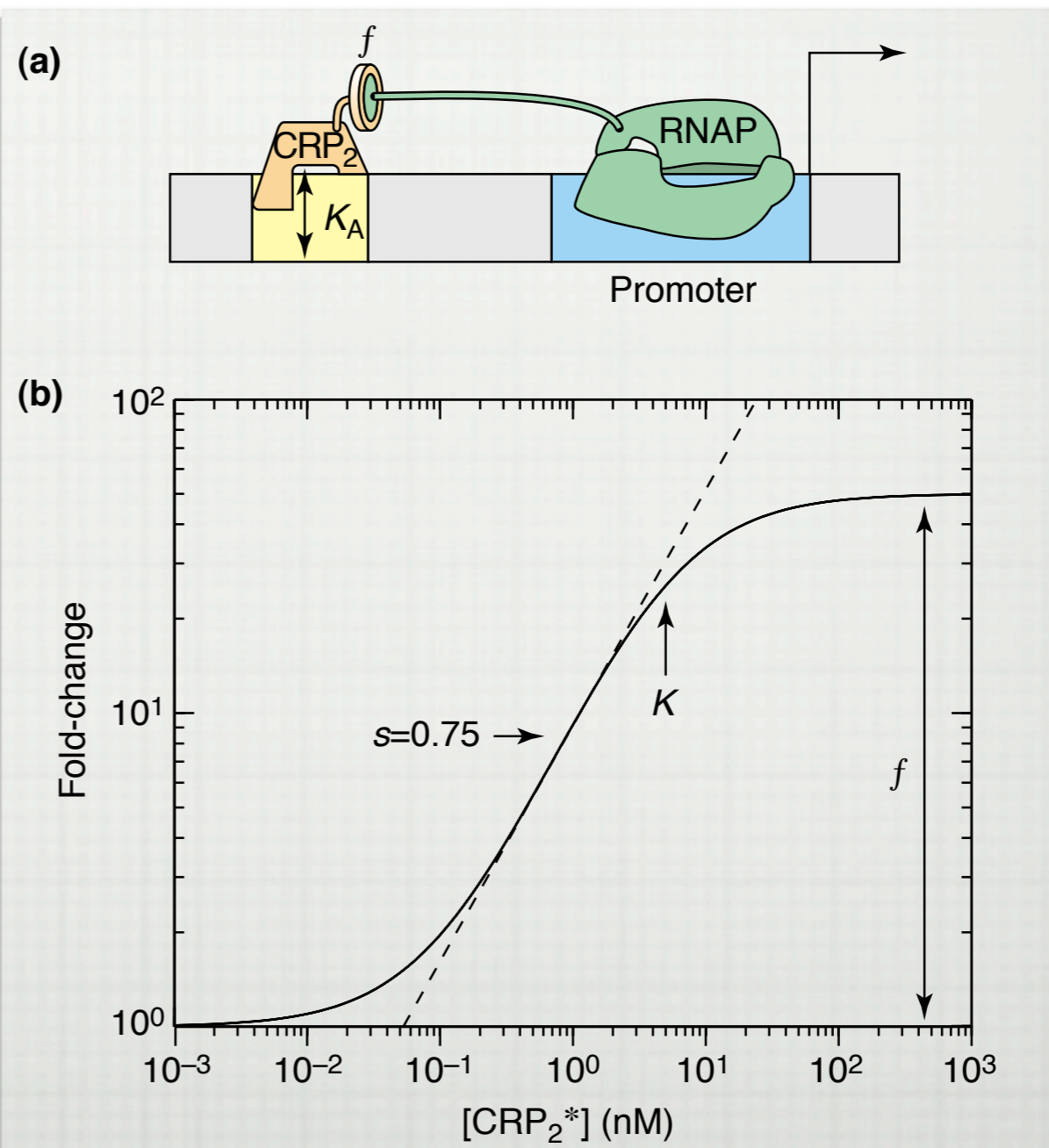
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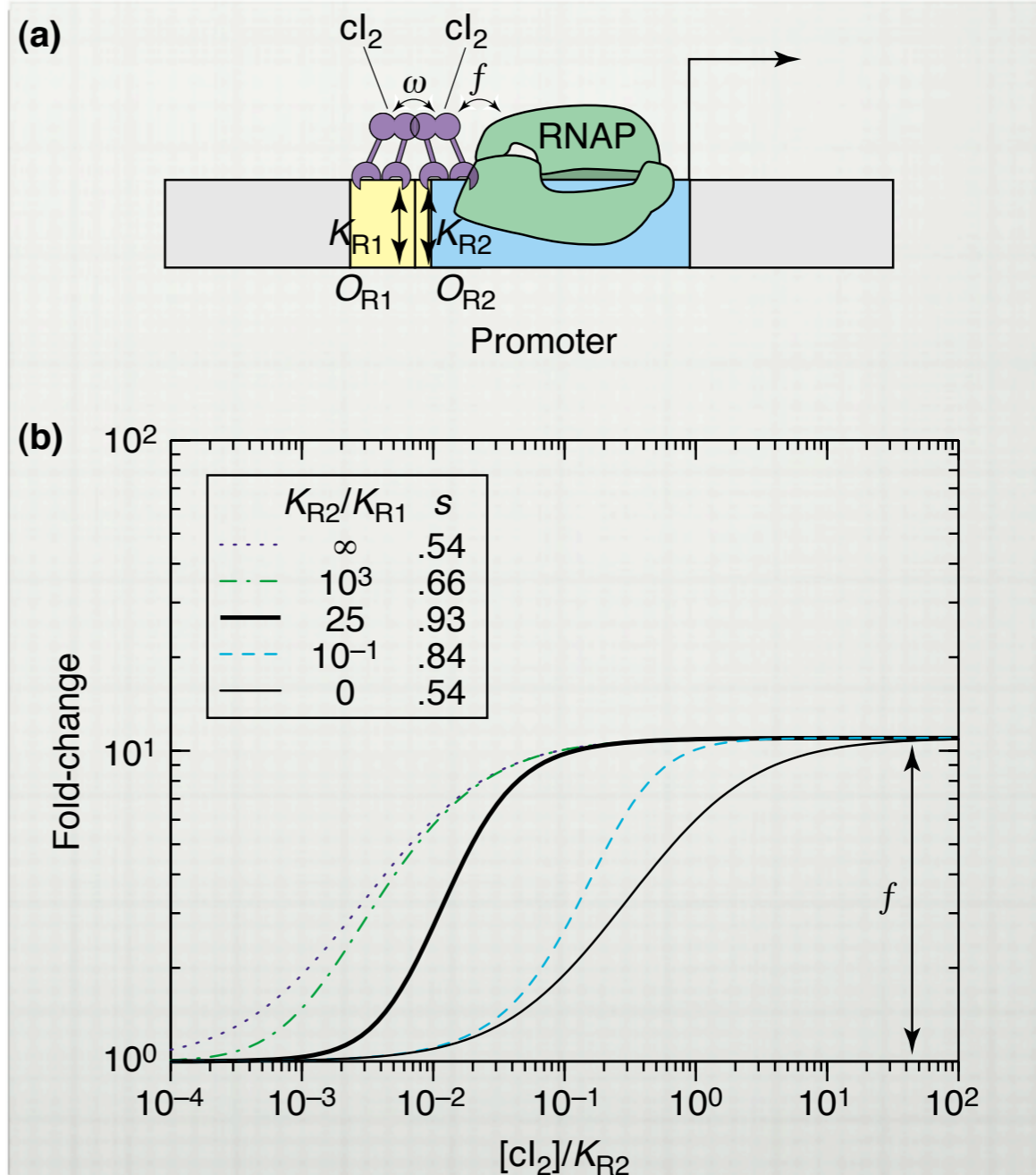


2. activation and helper: cl/P_{RM}

$$F = \frac{(1 + [H]/K_H + f[A]/K_A + f\Omega [H][A]/K_H K_A)}{(1 + [H]/K_H + [A]/K_A + \Omega [H]/K_H * [A]/K_A)}$$

log-log plot of F vs
 $[cl_2] = (\text{dimer}) \text{ TF}$
 for various values of K_{R2}/K_{R1}

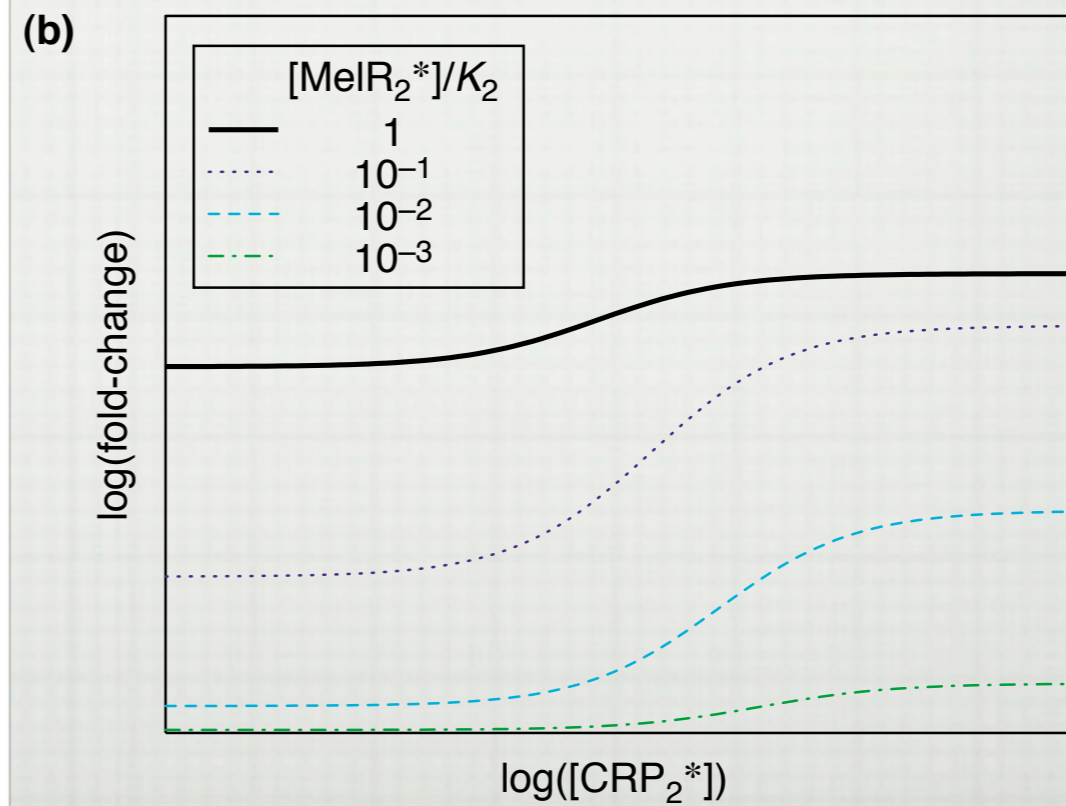
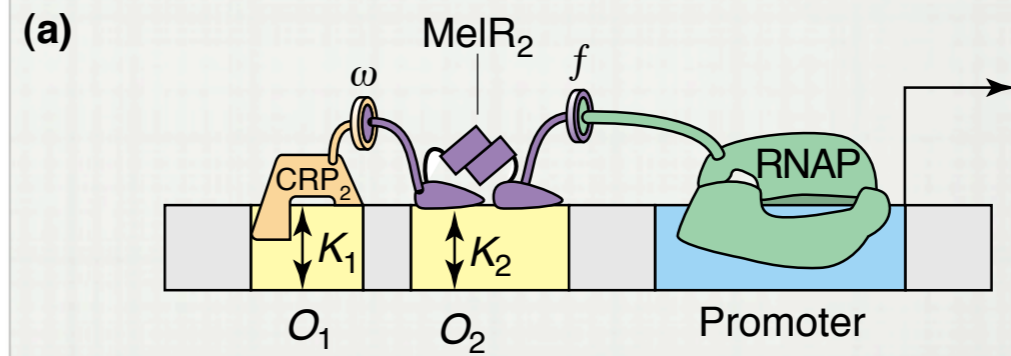
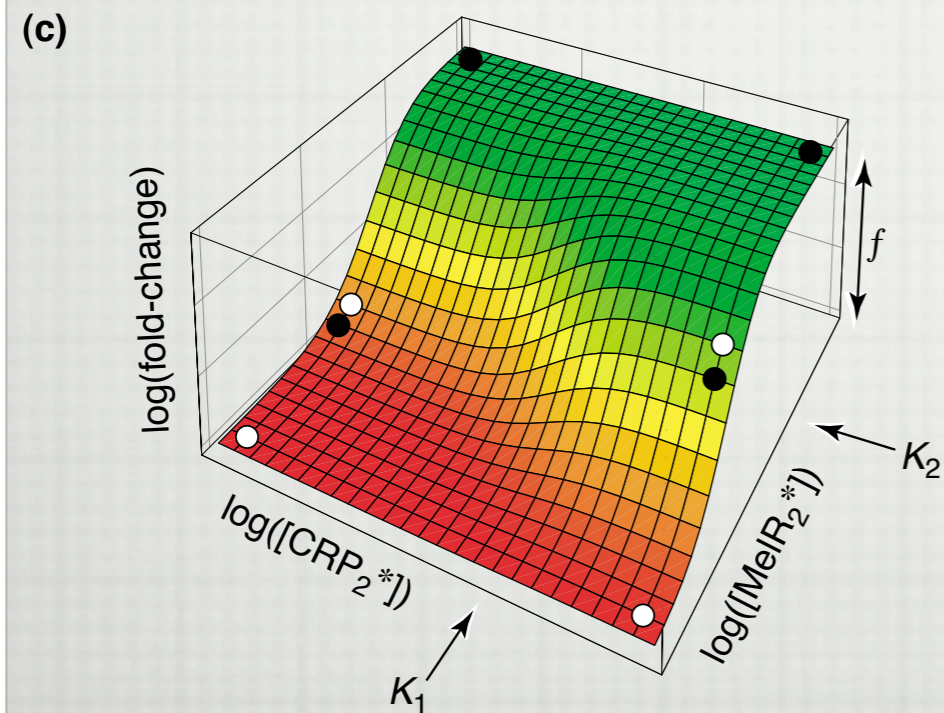
parameters: $K_{R2}, K_{R1}, f, \omega$
 sensitivity s



3. co-activation: CRP, MeIR₂/P_{RM}

F and params same as in 2.

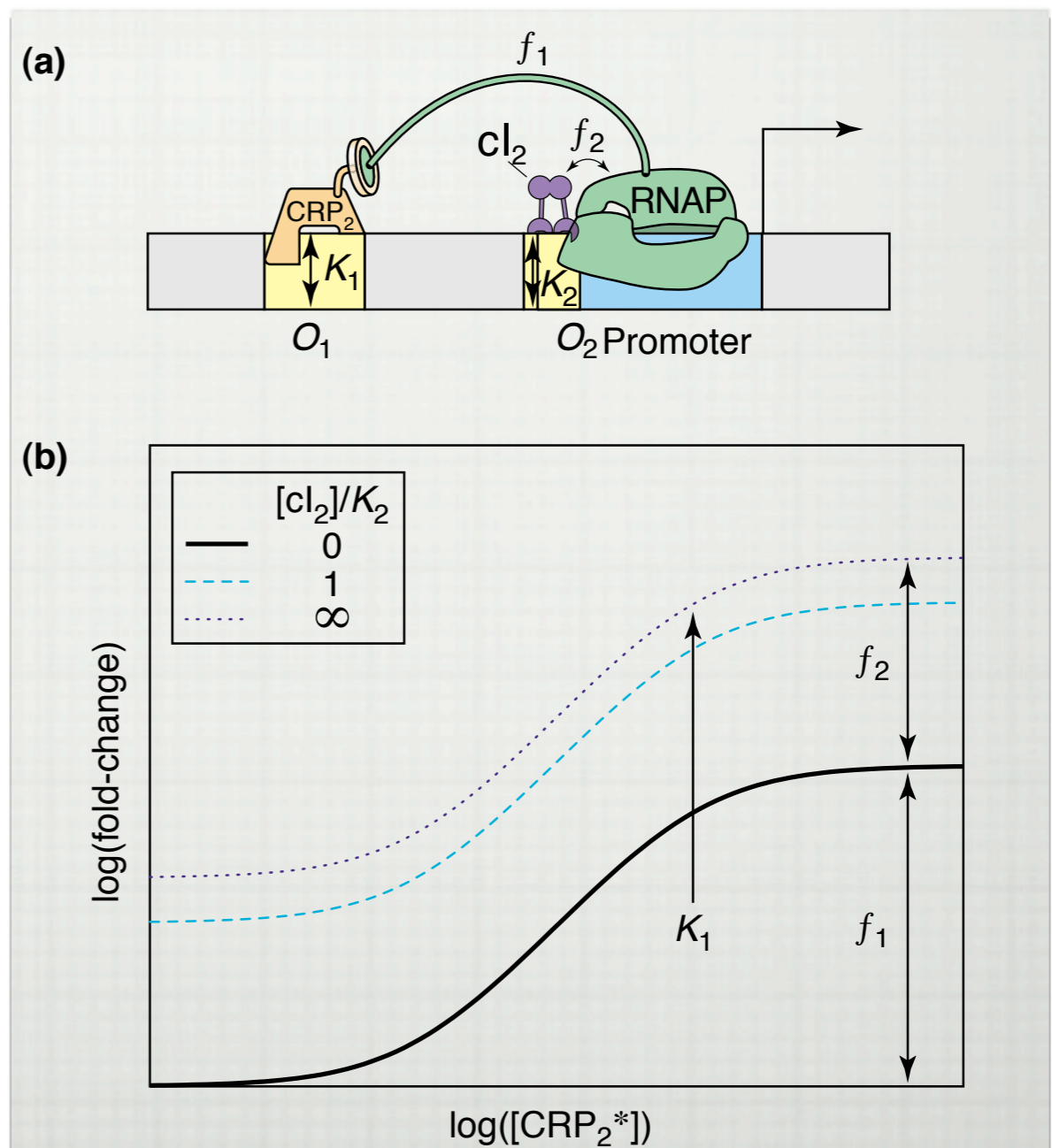
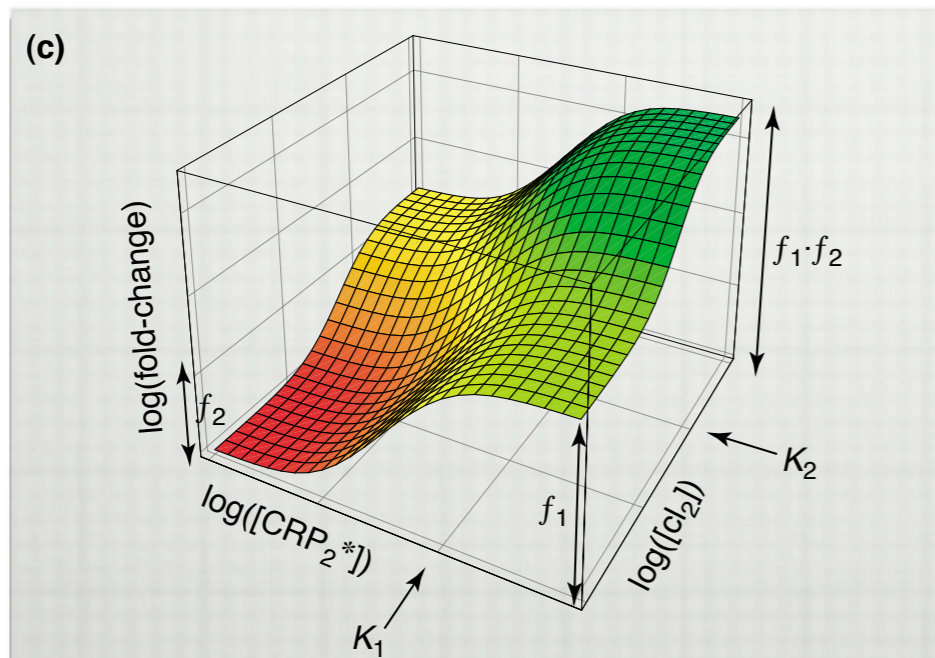
log-log plot of F vs
[C₁₂] and [MeIR₂]



4. dual activation: CRP, cI/P

$$F = \frac{(1 + f_1[A_1]/K_1 + f_2[A_2]/K_2 + \Omega f_1 f_2 [A_1][A_2]/K_1 K_2)}{(1 + [A_1]/K_1 + [A_2]/K_2 + \Omega [A_1][A_2]/K_1 K_2)}$$

here $\Omega=1$, no direct interaction between A_1 and A_2



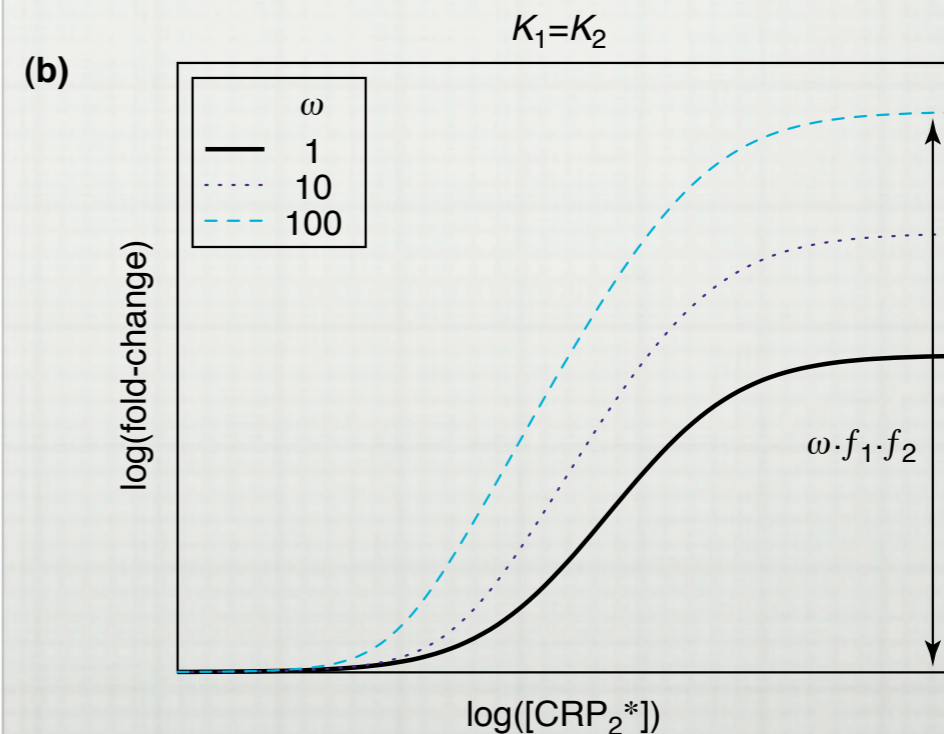
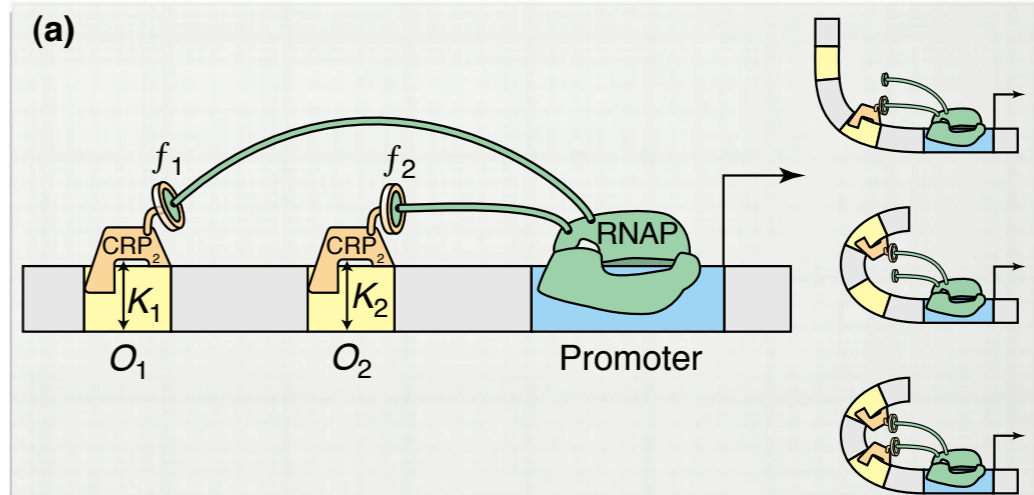
5. dual activation with direct cooperation: CRP, cI/P

CRP is known to activate at different distances

same F as in 4.

$$F = \frac{(1 + f_1[A_1]/K_1 + f_2[A_2]/K_2 + \Omega f_1 f_2 [A_1][A_2]/K_1 K_2)}{(1 + [A_1]/K_1 + [A_2]/K_2 + \Omega [A_1][A_2]/K_1 K_2)}$$

here $\Omega > 1$, there is direct interaction between A_1 and A_2 : looping DNA



next class ...

- various combinations of repressors
- thermo model for the derivation of the expressions for F

deliberately simple promoters

Elowitz et al. Mol Syst Biol. 2007 3:145

- The paper reports the synthesis of about 200 promoters glued to a reporter gene;
- the obtained DNA constructs can be seen as binary functions (most have 2 operators so 2 TF they can interact with).
- The constructs are simple promoter architectures, a priori no TF-TF contacts and no operator overlap.
- Constructs are classified in an original way as real-valued binary functions and then sequenced (why?)

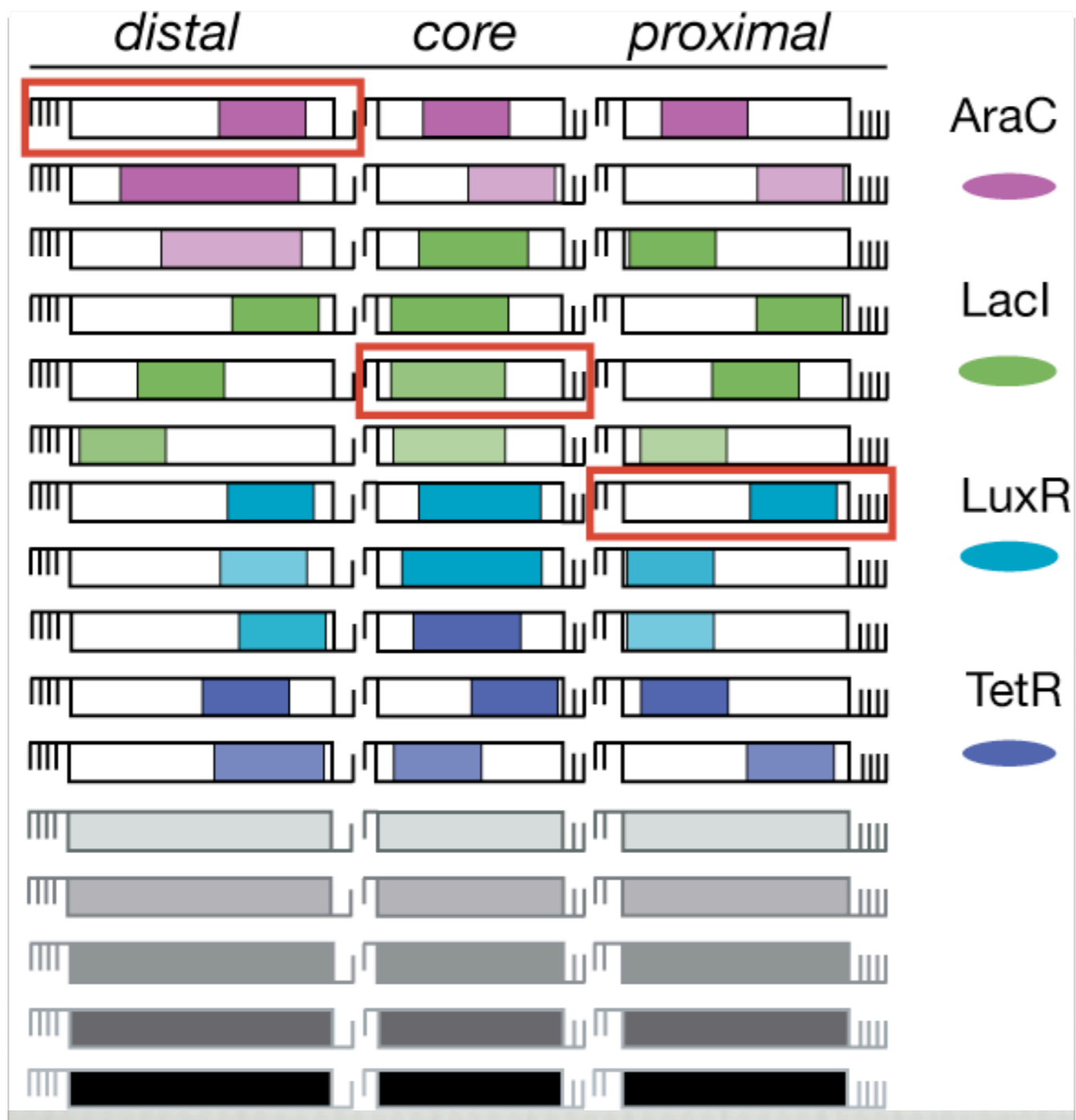
remarks

- A self-documenting automated bio-brick factory!
- yet output of a promoter::gene not a Boolean valued function of the concentrations of its TF/inputs (lac promoter has 4 output levels):
- could take "low" value of a few molecules per bacterium (1 nM)
- "high" value 1,000 molecules per bacterium (1M)

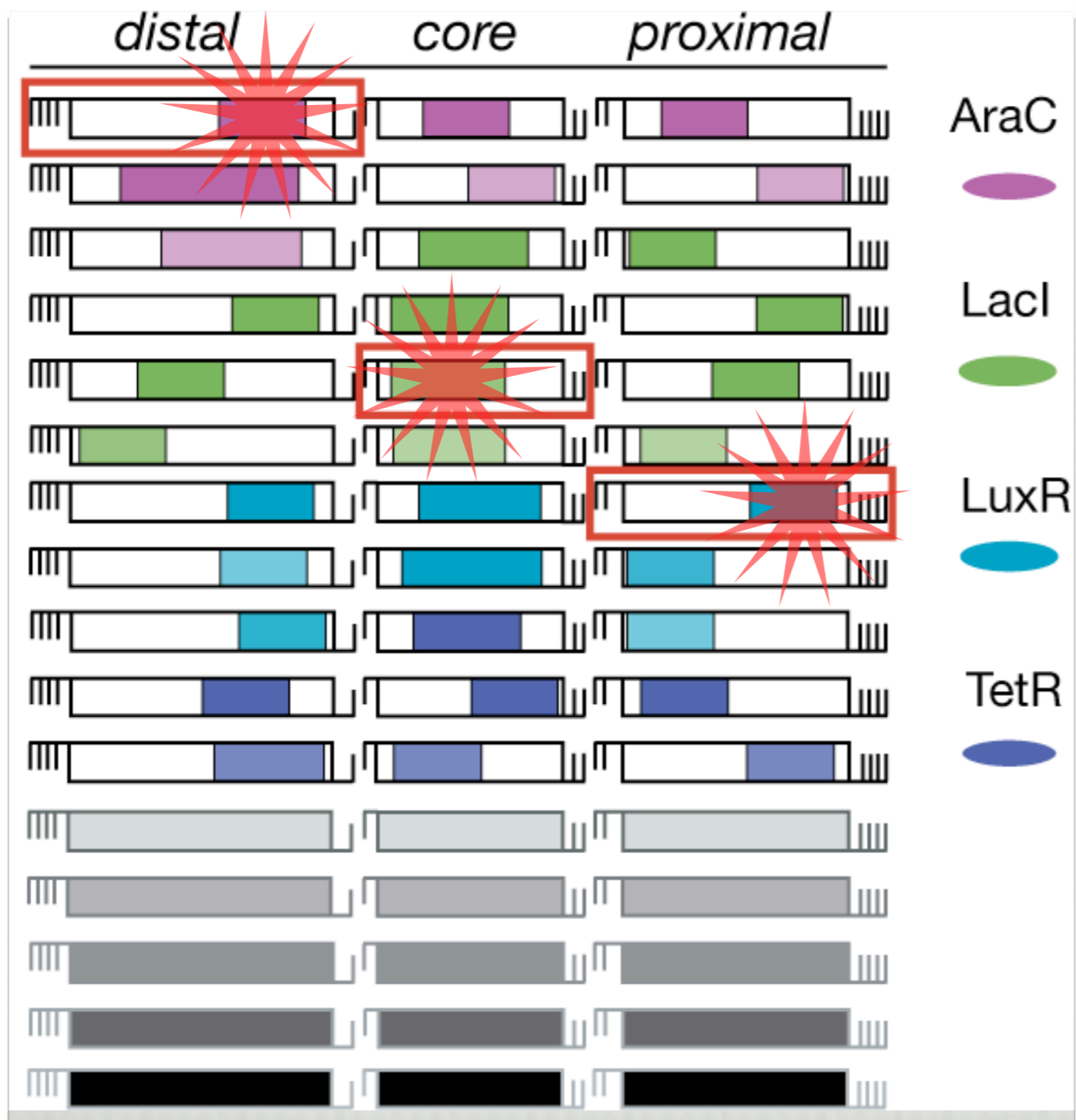
a combinatorial library of random promoter architectures

- 4096 of which 288 sequenced cassettes:
- 217 unique
- of which 27 binary (twofold response under 2 TFs)
- promoter = distal::core::proximal, device = promoter::G-luciferase
- TFs = Arac, LuxR (activators) -activated by Lara, VAI
- TetR, lacI (repressors) -inactivated by aTc, IPTG

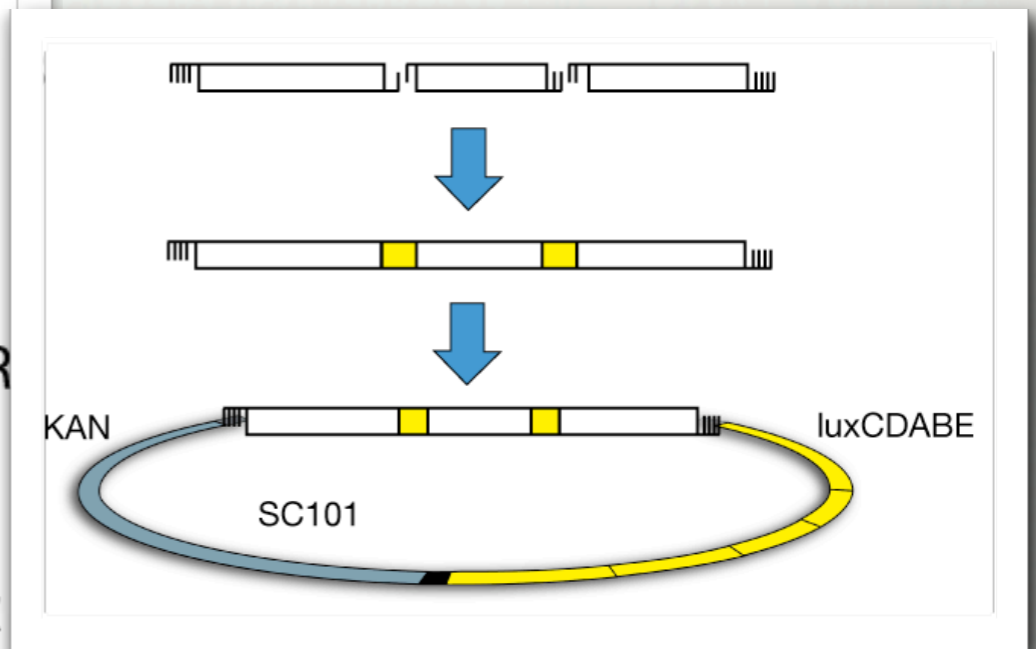
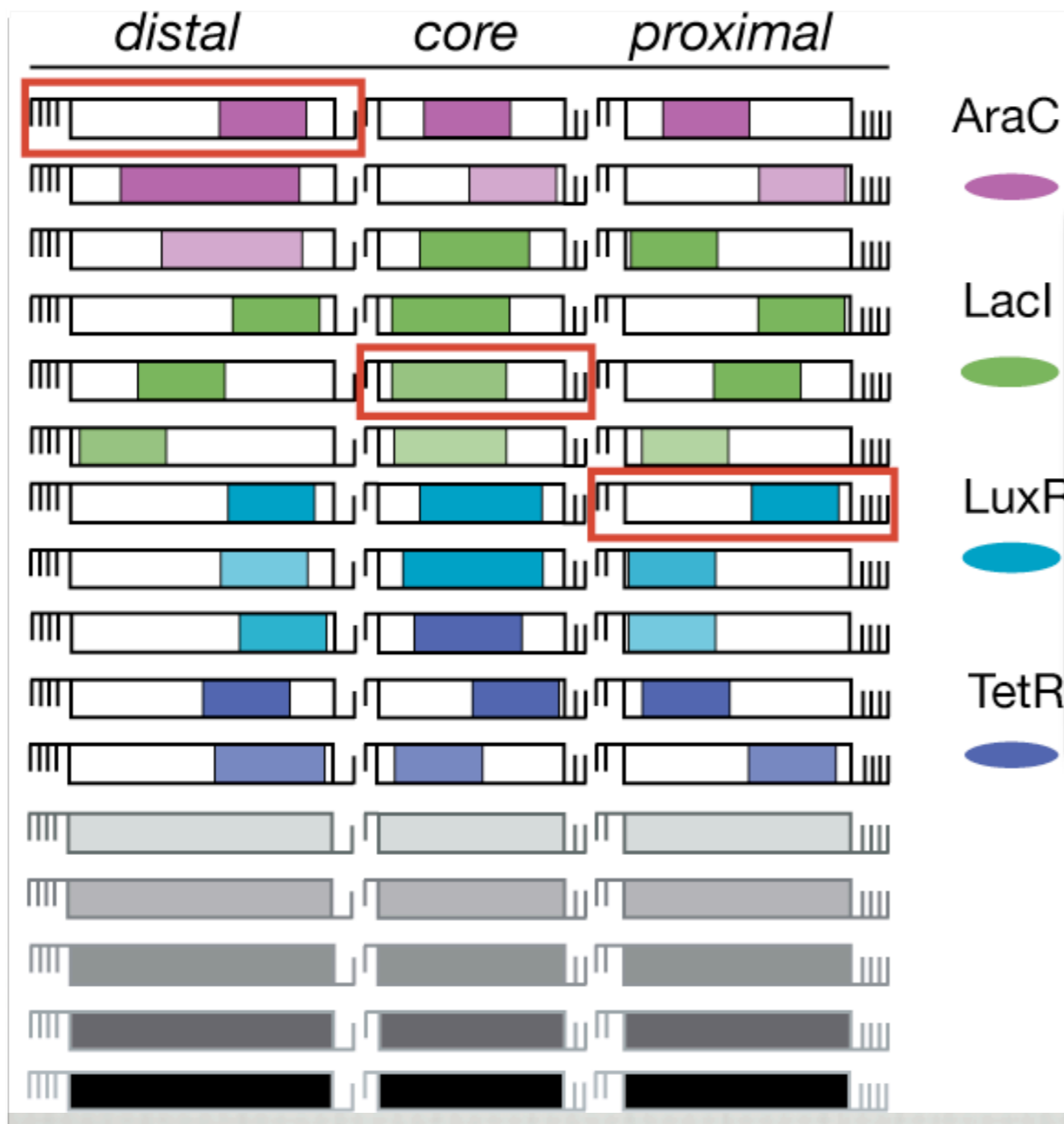
The library - sequence work



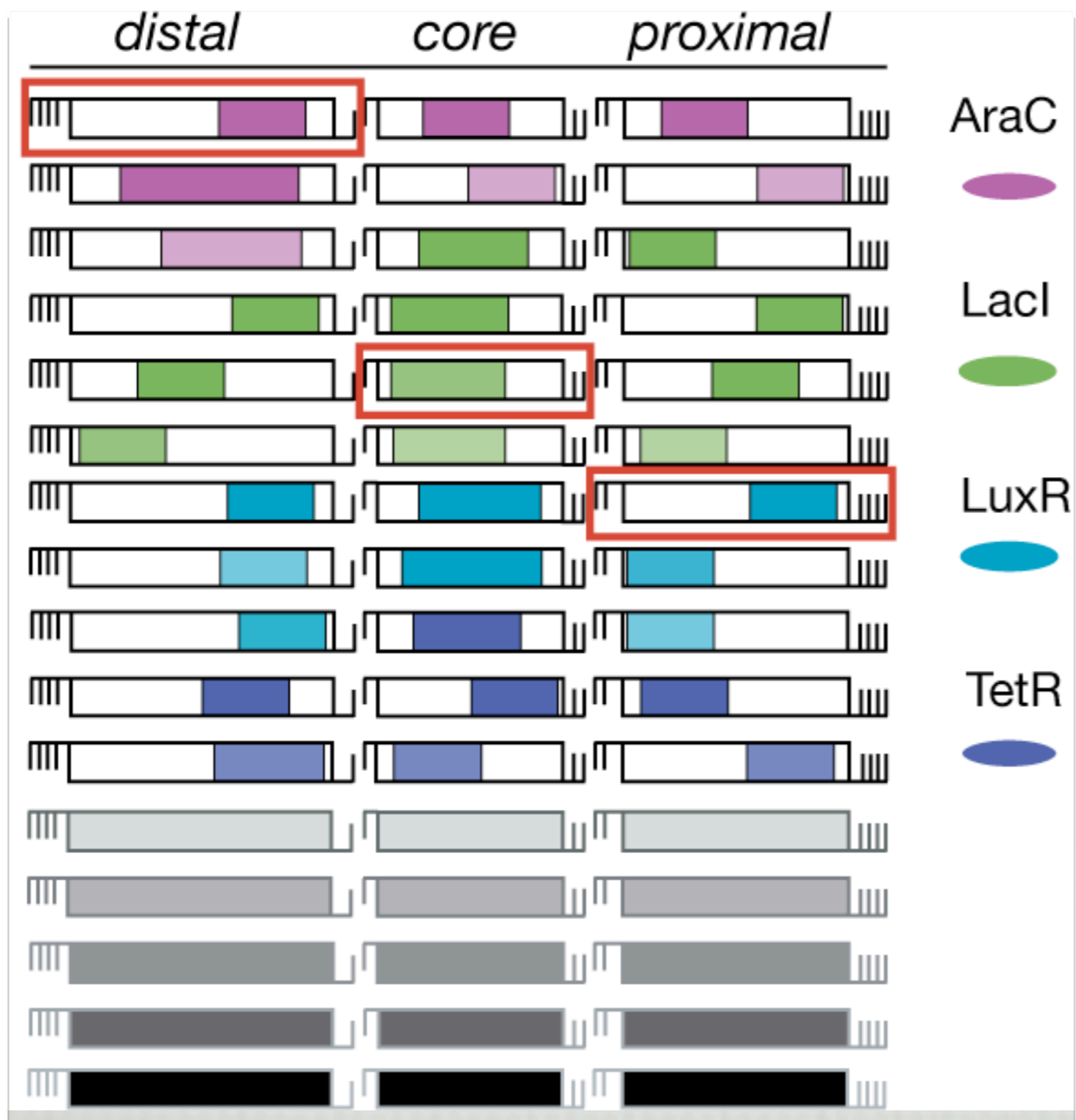
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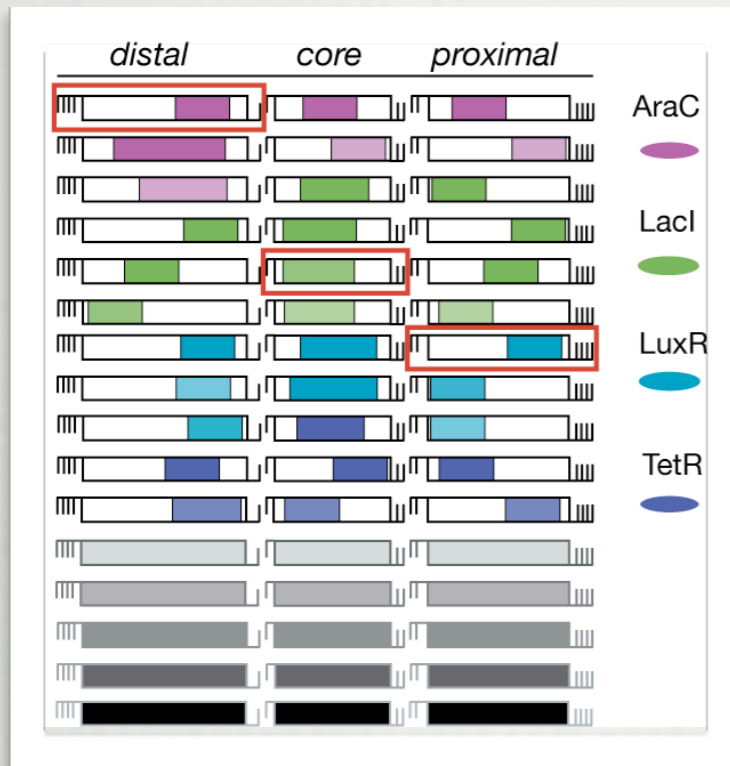
The library - sequence work



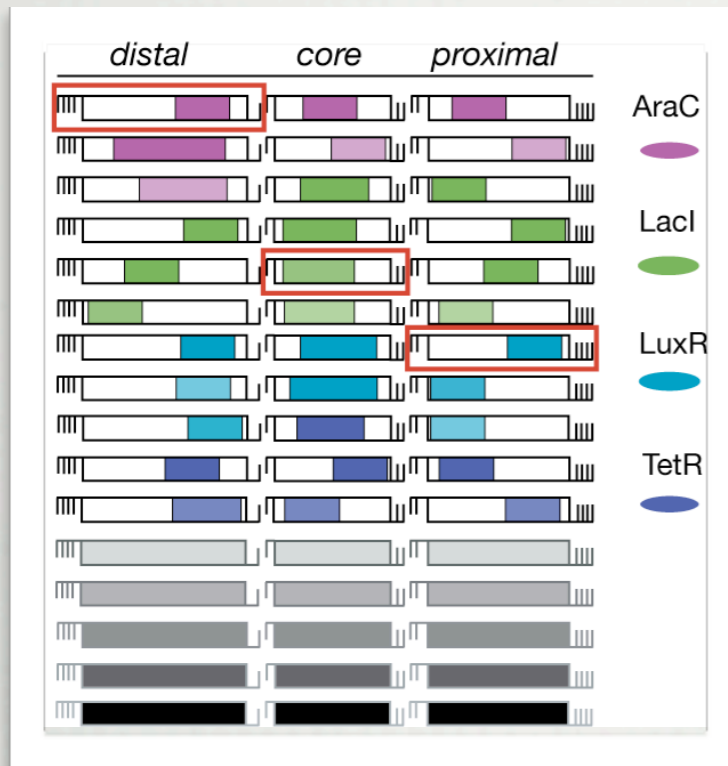
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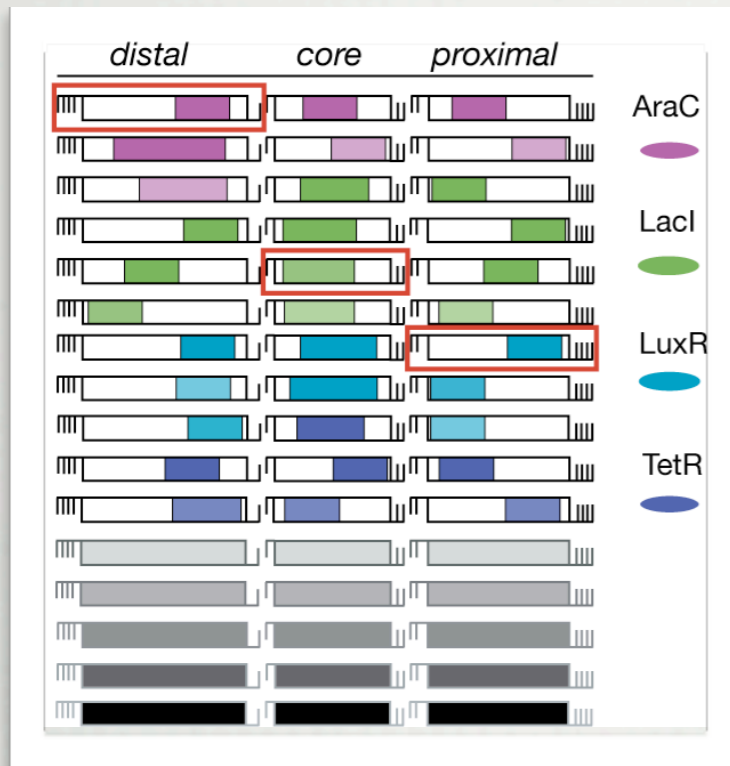
The library - sequence work



tcgag tacaacg tgcgtg ttagctgc cttt tagcaatt ttatcca **tagact** tgtgagc gctcacaatt **tataat** tcgtgcaat Ttttaaacctg taggatcgtacaggtg
 catg ttgcagcacaatcgacggaaaatcg ttaaaataggtat **ctga** cactcgcgagtg ttaa **atatta** agcacgttaAaaatttggacatcctagcatgtcca cctag

AraC I1 site -35 LacI Os site -10 +1 LuxR box

The library - sequence work



`tcgag`tacaacgtcgtgtagctgccttttagcaattttatcca**tagac**ttgtgagcgctcacaatt**tataat**tcgtgcaatTtttaa**acctgtaggatcgtacaggtg**
`catg`ttgcagcacaatcgacggaaatcgttaaaataggtat**ctg**aacactcgcgagtgtaa**atatta**agcacgttaAaaatt**ggacatcctagcatgtcca**cctag

AraC I1 site -35 LacI Os site -10 +1 LuxR box

288/4096 SEQUENCED

remarks

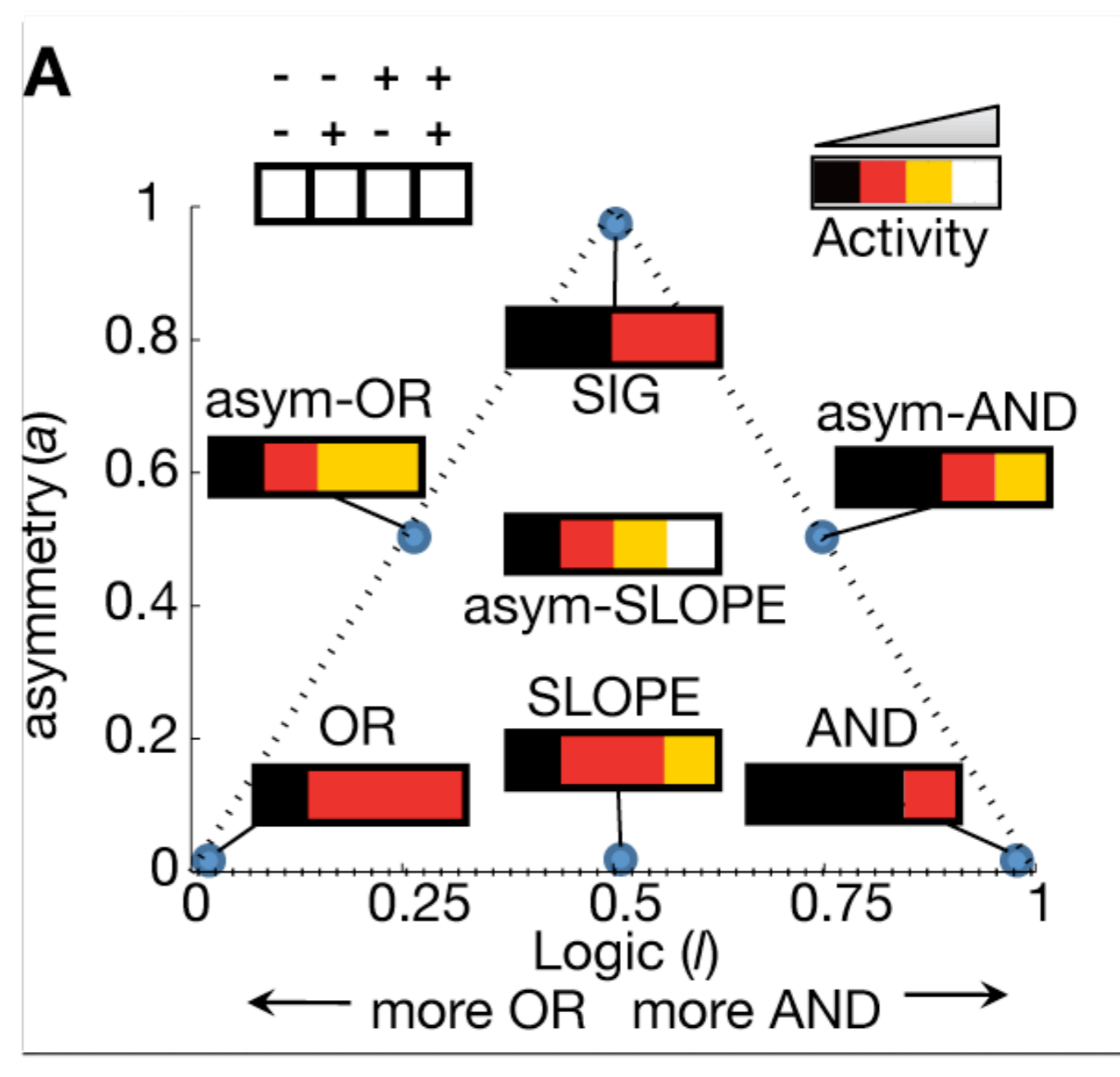
- The search is limited to neighbourhood of existing operators; it is really variation
- The observation is discretized (how robust is that?); who is listening to the outputs intervals;
- lacks a composition/impedance study; endogenousness?
- specificity, name space: possible to engineer chemical/TF specificity? wrt what is this complete?

typology (construction of the phenotype)

- regulatory range: exp-on/EXP-off [caveat: this is always >1 by def]
- logic type: from or $l=0$, to and $l=1$
- symmetry: from $a=0$ (complete symmetry) to $A=1$ (dependency in only 1 input) [works only for binary functions]

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typology (2)

- The level of TF is controlled indirectly by chemicals, and repressors are repressed, while activators are activated.
- So whatever the construct is, the attached function is monotonic increasing.
- The classification scheme -writing $b_1 < b_2 < b_3 < b_4$ for the increasing sequence of responses (by monotony b_1 , and b_4 are obtained for 00 and 11 inputs) - is:
 - the dynamic range $r = \log(b_4/b_1)$ in log scale
 - the asymmetry $a = \log(b_3/b_2)/r$ the b_3 to b_2 gap normalised to r so in 0 (fully symmetric) to 1 (unary function)
 - the and-ity $l = (\log(b_4) - 1/2(\log(b_3) + \log(b_2)))/r$ which is 0 if $b_4 = b_3 = b_2$, 1 (an OR) if $b_3 = b_2 = b_1$ (an AND)

model of RR promoter activity
under dual repression (Bintu)

- The r , a , and l trinity above can be defined in terms of the micro-trinity c_1 , c_2 , ω measuring the joint activity of a pair of repressors
- $P(R_1, R_2) = A / (c_1 R_1 + c_2 R_2 + \omega c_1 c_2 R_1 R_2)$
- A max promoter activity
- c_1 , c_2 TF efficiencies (at excluding RNAPol)
- $\omega = \text{cooperation} (>1)$

Looking for Mr Nice component

- computational models of transcription (eg "Transcriptional regulation by the numbers" Curr Opin Genet Dev -2005)
- evolution driven design (eg "Directed evolution of a genetic circuit" PNAS 2002)
- combinatorial approach (this paper)