

Why?

- Genome sequencing gives us new gene sequences
- Network biology gives us functional information on genes/proteins
- Analysis of mutants links unknown genes to diseases
- Can we learn anything from other known sequences about our new gene/protein?

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What is it?

ACCGGTATCCTAGGAC

ACC--TATCTTAGGAC

- Match the two sequences as closely as possible = aligned
- · Therefore, alignments need a score

















ACC**GG**TATCCTAGGAC

- Matches and substitutions are 'easy' to deal with.
 - We'll look at substitution matrices later.
- How do we score indels: gaps?

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How do we score gaps?

ACC**GG**TATCC---GAC

- A gap is a consecutive run of indels
- The gap length is the number of indels.
- The simple example here has two gaps of length 2 and 3

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How do we score gaps?

ACCGGTATCC---GAC

- Constant: Length independent weight
- Affine: Open and Extend weights.
- Convex: Each additional gap contributes less
- Arbitrary: Some arbitrary function on length

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Choosing Gap Penalties

- The choice of Gap Scoring Penalty is very sensitive to the context in which it is applied:
 - introns vs exons
 - protein coding regions
 - mis-matches in PCR primers

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Substitution Matrices

- Substitution matrices are used to score substitution events in alignments.
- Particularly important in Protein sequence alignments but relevant to DNA sequences as well.
- Each scoring matrix represents a particular theory of evolution

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Similarity/Distance

- Distance is a measure of the cost or replacing one residue with another.
- Similarity is a measure of how similar a replacement is.
- e.g. replacing a hydrophobic residue with a hydrophilic one.
- The logic behind both are the same and the scoring matrices are interchangeable.









How can we score a substitution in an aligned sequence?

 Amino acid property matrix
 Assign arbitrary values to the relatedness of different amino acids:
 e.g. hydrophobicity, charge, pH, shape, size





PAM Matrices

- · Ignore evolutionary direction
- Obtained frequencies for residue X being substituted by residue Y over time period Z
- Based on 1572 residue changes
- They defined a substitution matrix as 1 PAM (point accepted mutation) if the expected number of substitutions was 1% of the sequence length.

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PAM Matrices

To increase the distance, they multiplied the the PAM1 matrix.

PAM250 is one of the most commonly used.

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PAM - notes

The PAM matrices are rooted in the original datasets used to create the theoretical trees

They work well with closely related sequences

Based on data where substitutions are most likely to occur from single base changes in codons.

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PAM - notes

Biased towards conservative mutations in the DNA sequence (rather than amino acid substitutions) that have little effect on function/structure.

Replacement at any site in the sequence depends only on the amino acid at that site and the probability given by the table. This does not represent evolutionary processes correctly. Distantly related sequences usually have regions of high conservation (blocks).

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PAM - notes

36 residue pairs were not observed in the dataset used to create the original PAM matrix

A new version of PAM was created in 1992 using 59190 substitutions: Jones, Taylor and Thornton 1992 CAMBIOS 8 pp 275

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BLOSUM matrices

Henikoff and Henikoff 1991

Took sets of aligned ungapped regions from protein families from the BLOCKS database.

The BLOCKS database contain short protein sequences of high similarity clustered together. These are found by applying the MOTIF algorithm to the SWISS-PROT and other databases. The current release has 8656 Blocks.

BLOSUM matrices

Sequences were clustered whenever the %identify exceeded some percentage level.

Calculated the frequency of any two residues being aligned in one cluster also being aligned in another

Correcting for the size of each cluster.

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BLOSUM matrices

Resulted in the fraction of observed substitutions between any two residues over all observed substitutions.

The resulting matrices are numbered inversely from the PAM matrices so the BLOSUM50 matrix was based on clusters of sequence over 50% identity, and BLOSUM62 where the clusters were at least 62% identical.

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Summary so far...

- Gaps
 - Indel operations
 - Gap scoring methods
- Substitution matrices
 - DNA largely simple matrices
 - Protein matrices are based on probability
 - PAM and BLOSUM

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How do we do it?

- Like everything else there are several methods and choices of parameters
- The choice depends on the question being asked
 - What kind of alignment?
 - Which substitution matrix is appropriate?
 - What gap-penalty rules are appropriate?
 - Is a heuristic method good enough?

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• For proteins, using the affine gap penalty rule and a substitution matrix:

Query Length Matrix Gap (open/extend)

<35	PAM-30	9,1
35-50	PAM-70	10,1
50-85	BLOSUM-80	10,1
>85	BLOSUM-62	11,1

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Alignment Types

- Global: used to compare to similar sized sequences.
- Local: used to find similar subsequences.
- Ends Free: used to find joins/overlaps.

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Global Alignment

- Two sequences of similar length
- Finds the best alignment of the two sequences
- · Finds the score of that alignment
- Includes **ALL** bases from both sequences in the alignment and the score.
- · Needleman-Wunsch algorithm

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Needleman-Wunsch algorithm

- Gaps are inserted into, or at the ends of each sequence.
- The sequence length (bases+gaps) are identical for each sequence
- Every base or gap in each sequence is aligned with a base or a gap in the other sequence

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Needleman-Wunsch algorithm

- Consider 2 sequences S and T
- Sequence *S* has *n* elements
- Sequence *T* has *m* elements
- Gap penalty ?

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How do we score gaps?

ACC**GG**TATCC---GAC

- ACC--TATCT**TAG**GAC
- Constant: Length independent weight
- Affine: *Open* and *Extend* weights.
- Convex: Each additional gap contributes less
- Arbitrary: Some arbitrary function on length

 Lets score each gap as –1 times length

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Needleman-Wunsch algorithm

- Consider 2 sequences *S* and *T*
- Sequence *S* has *n* elements
- Sequence *T* has *m* elements
- Gap penalty –1 per base (arbitrary gap penalty)
- An alignment between base *i* in *S* and a gap in *T* is represented: (*S*_{*i*,-})
- The score for this is represented : $\sigma(S_i, -) = -1$



T -1 -1 2



Needleman-Wunsch algorithm

- Set up a array V of size n+1 by m+1
- Row 0 and Column 0 represent the cost of adding gaps to either sequence at the start of the alignment
- Calculate the rest of the cells row by row by finding the optimal route from the surrounding cells that represent a gap or match/mismatch
 This is easier to demonstrate than to explain

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 lets start by trying out a simple example alignment:

> S = ACCGGTATT = ACCTATC

































































Space Complexity

- To calculate each row we need the current row and the row above only.
- Therefore to get the score, we need $\mathrm{O}(\mathrm{n+m})$ space
- However, if we need the pointers as well, this increases to O(nm) space
- This is a problem for very long sequences – think about the size of whole genomes

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Global alignment in linear space

- Hirschberg 1977 applied a 'divide and conquer' algorithm to Global Alignment to solve the problem in linear space.
- Divide the problem into small manageable chunks
- The clever bit is finding the chunks

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Complexity

- After applying Hirschberg's divide and conquer approach we get the following:
 - Complexity O(mn)
 - Space O(min(m,n))
- For the proofs, see D.S. Hirschberg. (1977) Algorithms for the longest common subsequence problem. J. A.C.M 24: 664-667

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OK where are we?

- The Needleman-Wunsch algorithm finds the optimum alignment and the best score.
 NW is a dynamic programming algorithm
- Space complexity is a problem with NW
- Addressed by a divide and conquer algorithm
- What about local and ends-free alignments?



- Between two sequences, find the best two subsequences and their score.
- We want to ignore badly matched sequence
- Use the same types of substitution matrix and gap penalties
- Use a modification of the previous dynamic programming approach.

Smith-Waterman algorithm

- If S_i matches T_i then $\sigma(S_i, T_i) \ge 0$
- If they do not match or represent a gap then <=0
- Lowest allowable value of any cell is 0
- Find the cell with the highest value (*i*,*j*) and extend the alignment back to the first zero value
- The score of the alignment is the value in that cell
- A quick example if best...





















				r	ow					
		G	Т	Т	A	С	Т	G	Т	(S)
	0	0	0	0	0	0	0	0	0	
С	0	-1	-1	-1	-1	2	1	0	-1	
т	0	-1	1	1	0	1	4	3	2	
G	0	2	1	0	0	0	3	6	5	k
Т	0	1	4	3	2	1	2	5	8])
A	0	0	3	3	5	4	3	4	7	P
Т	0	-1	2	5	4	4	6	5	6	
С	0	0	1	4	4	6	5	5	5	

			ori	gin	and	d er	nd			
		G	Т	Т	A	С	Т	G	Т	(S)
	0	0	0	0	0	0	0	0	0	
С	0	-1	-1	-1	-1	2	1	0	-1	
Т	0	-1	1	1	0	1	4	3	2	
G	0	2	1	0	0	0	3	6	5	
Т	0	1	4	3	2	1	2	5	8	
A	0	0	3	3	5	4	3	4	7	
т	0	-1	2	5	4	4	6	5	6	
С	0	0	1	4	4	6	5	5	5	







- Dynamic programming algorithms can solve global, local and ends-free alignment
- · They give the optimum score and alignment using the parameters given
- Divide and conquer approaches make the space complexity manageable for smallmedium sized sequences



- We used a very simple gap penalty
- The Affine Gap penalty is most commonly used. - Cost to open a gap

Dynamic Programming Issues

- Cost to extend an open gap
- Need to track and evaluate the 'gap' state in the array





Real Life Sequence Alignment

- When searching multiple genomes, the sizes still get too big!
- Several approaches have been tried:
- Use huge parallel hardware:
 Distribute the problem over many CPUs
 Very expensive
- Implement in Hardware

 Cost of specialist boards is high
 Has been done for Smith-Waterman on SUN

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Real Life Sequence Alignment

- Use a Heuristic Method
 - Faster than 'exact' algorithms
 - Give an approximate solution
 - Software based therefore cheap
- Based on a number of assumptions:

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Assumptions for Heuristic Approaches

- Even linear time complexity is a problem for large genomes
- Databases can often be pre-processed to a degree
- Substitutions more likely than gaps
- Homologous sequences contain a lot of substitutions without gaps which can be used to help find start points in alignments

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Conclusions

- Dynamic programming algorithms are expensive but they give you the optimum alignment and exact score
- Choice of GAP penalty and substitution matrix are critically important
- Heuristic approaches are generally required for high throughput or very large alignments



Assumptions for Heuristic Approaches

- Even linear time complexity is a problem for large genomes
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- Substitutions more likely than gaps
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BLAST

Altschul, Gish, Miller, Myers and Lipman (1990) Basic local alignment search tool. J Mol Biol 215:403-410

- Developed on the ideas of FASTA
- Integrates the substitution matrix in the first stage of finding the *hot spots*
- Faster hot spot finding

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BLAST definitions

- Given two strings S_1 and S_2
- A segment pair is a pair of equal lengths substrings of S₁ and S₂ aligned without gaps
- A *locally maximal segment* is a segment whose alignment score (without gaps) cannot be improved by extending or shortening it.
- A *maximum segment pair (MSP)* in *S*₁ and *S*₂ is a segment pair with the maximum score over all segment pairs.

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BLAST Process

• Parameters:

- *w*: word length (substrings)
- *t*: threshold for selecting interesting alignment scores

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BLAST Process

• 1. Find all the *w*-length substrings from the database with an alignment score >*t*

- Each of these (similar to a hot spot in FASTA) is called a *hit*
- Does not have to be identical
- Scored using substitution matrix and score compared to
- the threshold *t* (which determines number found) - Words size can therefore be longer without losing
- sensitivity: AA 3-7 and DNA ~12

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BLAST Process

• 2. Extend hits:

- extend each hit to a local maximal segment
- extension of initial w size hit may increase or decrease the score
- terminate extension when a threshold is exceeded
- find the best ones (HSP)
- This first version of Blast did not allow gaps....

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(Improved) BLAST

Altshul, Madden, Schaffer, Zhang, Zhang, Miller & Lipman (1997) Gapped BLAST and PSI-BLAST:a new generation of protein database search programs. Nucleic Acids Research 25:3389-3402

- Improved algorithms allowing gaps
 - these have superceded the older version of BLAST
 - two versions: Gapped and PSI BLAST

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(Improved) BLAST Process

- · Find words or hot-spots
 - search each diagonal for two w length words such that score >=t
 - future expansion is restricted to just these initial words
 - we reduce the threshold t to allow more initial words to progress to the next stage

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(Improved) BLAST Process

- · Allow local alignments with gaps
 - allow the words to merge by introducing gaps
 - each new alignment comprises two words with a number of gaps
 - unlike FASTA does not restrict the search to a narrow band
 - as only two word hits are expanded this makes the new blast about 3x faster

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PSI-BLAST

- Iterative version of BLAST for searching for protein domains
 - Uses a dynamic substitution matrix
 - Start with a normal blast
 - Take the results and use these to 'tweak' the matrix
 - Re-run the blast search until no new matches occur
- Good for finding distantly related sequences but high frequency of false-positive hits

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BLAST Programs

blastp compares an amino acid query sequence against a protein sequence database.
 blastn compares a nucleotide query sequence against a nucleotide sequence database.
 blastx compares a nucleotide query sequence translated in all reading frames against a protein sequence database.
 tblastn compares a protein query sequence against a nucleotide sequence database.
 tblastn compares a protein query sequence against a nucleotide sequence database.
 tblastn compares a protein query sequence against a nucleotide sequence database.
 tblastn compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database. (SLOW)





Alignment Heuristics

- Dynamic Programming is better but too slow
- BLAST (and FASTA) based on several assumptions about good alignments

 substitutions more likely than gaps
 good alignments have runs of identical matches
- FASTA good for DNA sequences but slower
- BLAST better for amino acid sequences, pretty good for DNA, fastest, now dominant.